

Hemostatic Agents in Critically Ill Patients

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INTRODUCTION

Coagulation disorders and hemorrhage are common in critically ill patients due to myriad causes. These include blood loss, hemodilution, acquired platelet dysfunction, coagulation factor consumption in extracorporeal circuits, activation of fibrinolytic, fibrinogenolytic and inflammatory pathways, hypothermia, etc. Coagulation defects in critical patients may be either congenital or acquired. Acquired coagulopathy are often present in the critically ill as a result of prescribed oral anticoagulants (for example, warfarin, dabigatran, rivaroxaban, apixaban, edoxaban) and anti-platelet agents (for example: P2Y12 receptor inhibitors-clodogrel, prasugrel, ticagrelor). Hemorrhagic shock remains a leading cause of mortality in patients especially with trauma despite tremendous progress in acute care medicine.¹

Management of bleeding and coagulopathy should include identifying patients at risk, understanding the impact of various invasive interventions on hemostasis, institution of allogeneic blood and factor testing, utilizing point-of-care laboratory and understanding the limitations of monitoring techniques, concentrate-based therapies and use various hemostatic agents.¹ While treating coagulopathy, it has to be kept in mind that both hyper- and hypocoagulable states often coexist and hemostasis is a balance between these two.

To understand mechanism of various haemostatic agents, it is imperative to review the basic physiology of hemostasis. The process of hemostasis, is a complex and finely tuned interaction between various plasma proteins, platelets, blood flow, and the endothelium. Five components are crucial in the maintenance and regulation of hemostasis. These include: (1) endothelial cells; (2) platelets for plug formation; (3) coagulation factors for formation of insoluble fibrin clot; (4) coagulation inhibitors; and (5) fibrinolysis. The whole process can be categorized into:

- Primary hemostasis which is platelet aggregation and platelet plug formation
- Secondary hemostasis which is the deposition of insoluble fibrin.

Platelets are activated in a series of processes and the activated platelets adhere to the site of injury and to each other, forming a plug. Along with this, insoluble fibrin is generated by the coagulation cascade. This forms a mesh that is incorporated into and around the platelet plug. This mesh increases the strength and stabilizes the blood clot. These two processes act simultaneously and are mechanistically intertwined. The fibrinolysis pathway plays the role of "check point" in hemostasis regulating thrombus formation.^{2,3}

Hemostatic agents can be broadly divided into:

- Pharmacological systematic hemostatic agents
- Clotting factor concentrates.

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PHARMACOLOGICAL SYSTEMIC HEMOSTATIC AGENTS

Antifibrinolytic Agents

Antifibrinolytic drugs should ideally be used where there is hyperfibrinolysis, for example cardiopulmonary bypass, orthotopic liver transplantation, urological, orthopaedic operations, etc. Antifibrinolytic agents are lysine analog and protease inhibitors. Protease inhibitors for example aprotinin and nafamostat, inhibit serine proteases. Lysine analogs, for example tranexamic acid, ε-aminocaproic acid reversibly combine with plasminogen.⁴

Aprotinin

Aprotinin is a broad-spectrum protease inhibitors, reduces fibrinolysis and stabilizes platelet function. Multiple randomized control trial showed its efficacy in reducing blood loss and transfusion requirements in patients undergoing cardiothoracic, liver transplant and orthopedic surgeries.⁵⁻⁸ Aprotinin also has potential anti-inflammatory effect. This effect of aprotinin encouraged a more systemic use in infants undergoing cardiac surgery. However, several studies showed that use of aprotinin was associated with an increased risk of renal failure, myocardial infarction, heart failure, stroke, encephalopathy and mortality.^{9,10} Blood Conservation Using Antifibrinolytics in a Randomized Trial (BART) that looked into the effects of aprotinin, tranexamic acid and epsilon-aminocaproic acid on massive postoperative bleeding and death from any cause at 30 days, had to be terminated early because of a higher mortality in the aprotinin-treated patients.¹¹ This triggered suspension of manufacturing of aprotinin.

Nafamostat Mesilate

Nafamostat mesilate is synthetic protease inhibitor and inhibits thrombin, factors Xa and XIIa, kallikrein, plasmin and complement factors (C1r, C1s). It also works as an antifibrinolytic, anticoagulant and anti-inflammatory agent.¹² Clinically, it has been used in

the treatment of disseminated intravascular coagulation, acute pancreatitis and as a hemostatic agent in cardiac surgery. Nafamostat mesilate has a low-molecular weight and short half-life of only 8 minutes, which makes it suitable for use as an anticoagulant in extracorporeal circuits during CRRT in patients with a high risk of bleeding.^{12,13}

Tranexamic Acid

Tranexamic acid is in clinical practice since 1962. Tranexamic acid is a synthetic derivative of the amino acid lysine and binds the 5 lysine-binding sites on plasminogen. This inhibits plasmin formation and displaces plasminogen from the fibrin surface. It may also directly inhibit plasmin and partially inhibit fibrinolysis at higher concentrations.¹⁴ Tranexamic acid is usually given as a bolus dose of 10–15 mg per kg intravenous. As only a small fraction of administered tranexamic acid is metabolized and most is excreted unchanged by the kidney, the dose should be decreased in renal impairment.¹⁵ There are a number of clinical studies that tested its effectiveness in decreasing blood loss and transfusion requirement in surgical and nonsurgical settings. Various studies and meta-analysis in patients undergoing cardiopulmonary bypass, found that tranexamic acid was associated with a reduction in perioperative blood loss and allogenic blood transfusion requirements. However, timing of administration is crucial. It should be given preemptively before bleeding starts. Although there was an initial suspicion, but, tranexamic acid does not seem to be associated with increased risk of thrombotic phenomenon.^{16,17} Tranexamic acid has also been successfully used to reduce blood loss in orthopedic surgery particularly in knee arthroplasty and hip replacement.^{18,19} Studies evaluating the effect of high-dose tranexamic acid (10–40 mg per kg per hour) in primary orthotopic liver transplantation reported a significant reduction in intraoperative blood loss and transfusion requirements.^{20,21} The large multicentric CRASH 2 trial showed a significant reduction in all cause-mortality and bleeding in trauma patients receiving early tranexamic acid if administered over less than three hours of trauma. Another large multicentric trial (WOMAN) conducted in patients with postpartum hemorrhage showed a significant reduction in hemorrhage and need for surgery to treat severe bleeding. Potential side effects when using tranexamic acid, include increased risk of thromboembolic events and neurological side-effects. However, significant incidence of such serious side effect was not observed in those studies. Rapid intravenous administration of tranexamic acid may sometimes cause hypotension and should therefore be administered slowly as an infusion. Some literature reported the potential association of large doses of tranexamic acid with seizure activity in both adult and pediatric cardiac patients.^{22–24}

Epsilon-aminocaproic Acid

Epsilon-aminocaproic acid (EACA) is a competitive inhibitor of plasminogen activation and inhibits plasmin at higher doses. The usual recommended dose is 150 mg per kg as an intravenous bolus before surgery, followed by an infusion of 15 mg per kg per hour during the operation. EACA is largely eliminated unchanged by renal excretion. The terminal elimination half-life for aminocaproic acid is 1–2 hours. It is effective in several situations, such as prophylaxis of bleeding episodes in hemophiliacs, control of menorrhagia, gastrointestinal bleeding, obstetrical bleeding and in bleeding following cardiac and thoracic surgery. Major side effects from EACA include hypotension, cardiac arrhythmias, rhabdomyolysis and thromboembolic events.⁴ Several trials have studied the prophylactic administration of EACA in patients

undergoing cardiopulmonary bypass. They found that EACA may be useful in reducing blood loss and transfusion. EACA has been studied in noncardiac surgery as well.^{25,26} A meta-analysis of three studies including 1691 patient undergoing total knee replacement showed that EACA is as effective as tranexamic acid in reducing estimated blood loss and transfusion.²⁷ EACA has also been studied in patients undergoing orthotopic liver transplantation. Data from 1170 consecutive transplants patients showed that EACA decreased intraoperative blood loss and showed a trend toward improved graft and patient survival.²⁸

Vasopressin Analogue: Desmopressin

Desmopressin acetate is pharmacologically altered form of naturally occurring vasopressin by deamination of hemicysteine at position 1 and substitution of D-arginine for L-arginine at position 8. Beside antidiuretic action, desmopressin causes the endothelial release of factor VIII and von Willebrand factor into the plasma. The released factors form a complex with platelets and enhance their ability to aggregate. Desmopressin at a dose of 0.3 mg per kg has been used in hemophilia A, von Willebrand's disease, uremic thrombocytopenia, and in perioperative settings. These doses can be repeated at intervals of 12–24 hours, but tachyphylaxis may occur. Side effects correspond with its antidiuretic and vasomotor effects. Hyponatremia, hypertension, tachycardia, nausea, malaise, headache, fatigue, flushing, dizziness are some of the common side effects with desmopressin.^{15,29} Few initial studies showed usefulness of desmopressin in patients pretreated with aspirin and undergoing cardiac surgery. But a meta-analysis of 72 trials found that the use of desmopressin in cardiac surgery, resulted in a small decrease in perioperative blood loss, but was not associated with a beneficial effect on other clinical outcomes.^{17,30,31} Desmopressin was also tried in several noncardiac surgeries for example spine and orthopedic surgeries which yielded mixed results.^{32,33} One of most common agent used in uremic patients with active bleeding is desmopressin. Desmopressin doses for uremic bleeding range from 0.3 µg/kg to 0.4 µg/kg intravenously or subcutaneously. DDAVP administration should not be repeated because of the risk of tachyphylaxis. It is postulated that tachyphylaxis occurs as a result of depletion of factor VIII and vWF endothelial stores.³⁴

Estrogens

Estrogen has also been used in uremic bleeding by virtue of its ability to decrease production of I-arginine, which is a precursor of NO. By decreasing production of cGMP, it increases production of thromboxane A2 and ADP. These are crucial contributors to formation of platelet plugs, decrease of antithrombin III and protein S levels, and increase factor VII concentrations. Various studies showed estrogen can decrease bleeding time.^{35,36}

Ethamsylate

Ethamsylate acts by reducing thromboxane A2 and prostacyclin biosynthesis and improving platelet homo- and heterotypic adhesiveness. Ethamsylate has been used in dysfunctional uterine bleeding, periventricular hemorrhage in very low-birth weight babies, perioperative scenarios.^{37,38}

Clotting Factor Concentrates

Fresh Frozen Plasma

Fresh frozen plasma is the fluid portion of a unit of whole blood that is frozen usually within 8 hours. FFP contains all clotting factors, fibrinogen, albumin, protein C, protein S, antithrombin,

tissue factor pathway inhibitor. Dose of FFP is 10–20 mL/kg which raises factor levels by approximately 20% and that is enough to maintain hemostasis. FFP should be ABO compatible. FFP is used for a planned invasive procedure in the presence of abnormal coagulation tests, reversal of warfarin in the presence of active bleeding or for planned procedure when vitamin K is inadequate to reverse the warfarin effect, during plasma exchange, congenital or acquired factor deficiency with no alternative therapy and as part of massive transfusion protocol.³⁹

Prothombin Complex Concentrate

Prothombin complex concentrate (PCC) may be either three-factor (i.e., factors II, IX and X) or four-factor (i.e., factors II, VII, IX and X) concentrates with a concentration approximately 25 times higher than in normal plasma.⁴⁰ Indication for PCC are urgent reversal of warfarin, congenital or acquired deficiency of vitamin K-dependent clotting factors and haemophilia B. Various studies compared efficacy of FFP to that of PCC. A systematic review of 14 studies found that PCC is more effective in reversing warfarin and decreasing INR.⁴¹ Dose of PCC is 1–2 mL/kg which is much lower than FFP. Another advantage of PCC over FFP is that PCC does not cause transfusion associated acute lung injury (TRALI).⁴²

Recombinant Factor VIIa

Recombinant factor VIIa is produced by transfection of the human factor VII gene into baby hamster kidney cells cultured in bovine albumin. It has an amino acid sequence identical to that of plasma-derived factor VII. rFVIIa binds to the surface of activated platelets and promotes factor X activation and thrombin generation localized at the site of injury without widespread thrombosis. This activation is independent of TF. Standard doses are 90–120 µg/kg given every 2–3 hours till effective hemostasis is achieved.^{43,44}

FDA approved uses of rFVIIa are:

- Treatment of bleeding episodes in hemophilia A or B patients with inhibitors to factor VIII or factor IX and in patients with acquired hemophilia
- Prevention of bleeding in surgical interventions or invasive procedures in hemophilia A or B patients with inhibitors to factor VIII or factor IX and in patients with acquired hemophilia
- Treatment of bleeding episodes in patients with congenital FVII deficiency
- Prevention of bleeding in surgical interventions or invasive procedures in patients with congenital FVII deficiency

Apart from these, other off-label uses of rFVIIa are spontaneous intracranial hemorrhage, trauma, postpartum hemorrhage, cardiac surgery, liver surgery etc.^{43,45}

A recent systematic review and meta-analysis of 12 studies including 1244 patients with acquired hemophilia found that rFVIIa is effective in bleeding control with good safety profile.⁴⁶ Another meta-analysis evaluated its efficacy and safety for treatment of bleeding in major abdominal, urological and vascular surgery. The study concluded that rFVIIa achieved at least a reduction of bleeding and that the probability of survival was increased in patients responding to rFVIIa. rFVIIa was not associated with an increased risk of thromboembolism compared with placebo.⁴⁷ A randomized control trial of 841 patients with intracerebral hemorrhage showed that rFVIIa at dose of 80 µg per kg reduced expansion of the hematoma but did not improve survival or functional outcome after intra cerebral hemorrhage.⁴⁸

The appropriate management of patients with bleeding and coagulopathy remains a major challenge in routine clinical practice. A multidisciplinary approach and adherence to evidence-based guidance are key to improving patient outcomes. Selection of the most appropriate haemostatic pharmacological agents, requires not only consideration of the clinical evidence supporting the efficacy of the various agents, but also the available safety data to ensure that the benefits of this approach are not jeopardized by the risk. Correction of hypothermia, hypocalcemia, metabolic acidosis should precede use of pharmacological hemostatic agents. Dynamic parameters of coagulation may be more useful than static parameter in guiding management. Most studies on hemostatic agents have been carried out in trauma and perioperative settings. Further investigations are required to evaluate efficacy and safety of these hemostatic agents in more complex, critically care settings.

REFERENCES

1. Ghadimi K, Levy JH, Welsby JJ. Perioperative management of the bleeding patient. *Br J Anaesth.* 2016;117(suppl 3):iii18–iii30.
2. Zaidi A, Green L. Physiology of haemostasis, Anaesthesia and intensive care medicine 2019, [Article in press].
3. Gale AJ. Continuing education course #2: current understanding of hemostasis. *Toxicol Pathol.* 2011;39:273–280.
4. Porte RJ, Leebeek FWG. Pharmacological strategies to decrease transfusion requirements in patients undergoing surgery *Drugs* 2002;62:2193–2211.
5. Levy JH. Efficacy and safety of aprotinin in cardiac surgery *Orthopedics* 2004;27:S659–662.
6. Samama CM. A direct antifibrinolytic agent in major orthopedic surgery. *Orthopedics* 2004;27:S675–680.
7. Bitan FD. Aprotinin in spine surgery: review of the literature. *Orthopedics* 2004;27:S681–683.
8. Molenaar IQ, Warnaar N, Groen H, Tenvergert EM, Slooff MJ, Porte RJ. Efficacy and safety of antifibrinolytic drugs in liver transplantation: a systematic review and meta-analysis. *Am J Transplant* 2007;7:185–194.
9. Mangano DT, Tudor IC, Dietzel C. The risk associated with aprotinin in cardiac surgery. *N Engl J Med* 2006;354:353–365.
10. Karkouti K, Beattie WS, Dattilo KM, McCluskey SA, Ghannam M, Hamdy A, et al. A propensity score case-control comparison of aprotinin and tranexamic acid in high-transfusion-risk cardiac surgery. *Transfusion* 2006;46:327–338.
11. Fergusson DA, Hébert PC, Mazer CD, Fremes S, MacAdams C, Murkin JM, et al. A comparison of aprotinin and lysine analogues in high-risk cardiac surgery. *N Engl J Med* 2008;358:2319–2331.
12. Mahdy AM, Webster NR. Perioperative systemic haemostatic agents. *Br J Anaesth.* 2004;93:842–858.
13. Choi, Ji-Young, Kang, Yun-Jeong, Jang, Hye Min, Jung, Hee-Yeon; Cho, Jang-Hee; Park, Sun-Hee, Kim, Yong-Lim, Kim, Chan-Duck. Nafamostat Mesilate as an Anticoagulant During Continuous Renal Replacement Therapy in Patients With High Bleeding Risk: A Randomized Clinical Trial. *Medicine.* 2015;94(52):p e2392. doi: 10.1097/MD.0000000000002392.
14. Verstraete M. Clinical application of inhibitors of fibrinolysis. *Drugs* 1985;29:236–261.
15. Franck M, Sladen RN. Drugs to prevent and reverse anticoagulation. *Anesthesiol Clin North America* 1999;17:799–811.
16. Laupacis A, Fergusson D. Drugs to minimize perioperative blood loss in cardiac surgery: meta-analyses using perioperative blood transfusion as the outcome. The International Study of Perioperative Transfusion (ISPOT) Investigators. *Anesth Analg* 1997;85:1258–1267.
17. Levi M, Cromheecke ME, de Jonge E, Prins MH, de Mol BJ, Briët E, et al. Pharmacological strategies to decrease excessive blood loss in cardiac surgery: a metaanalysis of clinically relevant endpoints. *Lancet* 1999;354:1940–1947.



18. Hiippala ST, Strid LJ, Wennerstrand MI, Arvela JV, Niemelä HM, Mäntylä SK, et al. Tranexamic acid radically decreases blood loss and transfusions associated with total knee arthroplasty. *Anesth Analg* 1997;84:839–844.
19. Ekbäck G, Axelsson K, Rytberg L, Edlund B, Kjellberg J, Weckström J, et al. Tranexamic acid reduces blood loss in total hip replacement surgery. *Anesth Analg* 2000;91:1124–1130.
20. Boylan JF, Klinck JR, Sandler AN, Arellano R, Greig PD, Nierenberg H, et al. Tranexamic acid reduces blood loss, transfusion requirements, and coagulation factor use in primary orthotopic liver transplantation. *Anesthesiology* 1996;85:1043–1048.
21. Dalmau A, Sabaté A, Acosta F, Garcia-Huete L, Koo M, Sansano T, et al. Tranexamic acid reduces red cell transfusion better than epsilon-aminocaproic acid or placebo in liver transplantation. *Anesth Analg* 2000;91:29–34.
22. The WOMAN Trial Collaborators. Effect of early tranexamic acid administration on mortality, hysterectomy, and other morbidities in women with post-partum haemorrhage (WOMAN): an international, randomised, double-blind, placebo-controlled trial. *Lancet*. 2017;389:2105–2116.
23. Roberts I, Shakur H, Coats T, Hunt B, Balogun E, Barnetson L, et al. The CRASH-2 trial: a randomised controlled trial and economic evaluation of the effects of tranexamic acid on death, vascular occlusive events and transfusion requirement in bleeding trauma patients. *Health Technol Assess*. 2013;17:1–79.
24. Seizures after cardiac surgery. Hunter GR, Young GB *J Cardiothorac Vasc Anesth*. 2011;25:299–305.
25. Leff J, Rhee A, Nair S, Lazar D, Sathyanarayana SK, Shore-Lesserson L. A randomized, double-blinded trial comparing the effectiveness of tranexamic acid and epsilon-aminocaproic acid in reducing bleeding and transfusion in cardiac surgery. *Ann Card Anaesth*. 2019;22:265–272.
26. Blaine KP, Press C, Lau K, Sliwa J, Rao VK, Hill. Comparative effectiveness of epsilon-aminocaproic acid and tranexamic acid on postoperative bleeding following cardiac surgery during a national medication shortage. *J Clin Anesth*. 2016;35:516–523.
27. Riaz O, Aqil A, Asmar S, Vanker R, Hahnel J, Brew C, et al. Epsilon-aminocaproic acid versus tranexamic acid in total knee arthroplasty: a meta-analysis study. *J Orthop Traumatol*. 2019;20:28.
28. Mangus RS, Kinsella SB, Fridell JA, Kubal CA, Lahsaei P, Mark LO, et al. Aminocaproic Acid (amicar) as an alternative to aprotinin (trasylo) in liver transplantation. *Transplant Proc*. 2014;46:1393–1399.
29. Stoof SC, Cnossen MH, de Maat MP, Leebeek FW, Kruip MJ. Side effects of desmopressin in patients with bleeding disorders Haemophilia. 2016;22:39–45.
30. Dilthey G, Dietrich W, Spannagl M, Richter JA. Influence of desmopressin acetate on homologous blood requirements in cardiac surgical patients pretreated with aspirin. *J Cardiothorac Vasc Anesth* 1993;7:425–430.
31. Sheridan DP, Card RT, Pinilla JC, Harding SM, Thomson DJ, Gauthier L, et al. Use of desmopressin acetate to reduce blood transfusion requirements during cardiac surgery in patients with acetylsalicylic-acid-induced platelet dysfunction. *Can J Surg* 1994;37:33–36.
32. Guay J, Reinberg C, Rivard GE, Poitras B, Mathews S, David M. DDAVP does not reduce bleeding during spinal fusion for idiopathic scoliosis. *Can J Anaesth* 1990;37:514.
33. Flordal PA, Ljungstrom KG, Ekman B, Neander G. Effects of desmopressin on blood loss in hip arthroplasty. Controlled study in 50 patients. *Acta Orthop Scand* 1992;63:381–385.
34. Hedges SJ, Dehoney SB, Hooper JS, Amanzadeh J, Busti AJ. Evidence-based treatment recommendations for uremic bleeding. *Nat Clin Pract Nephrol*. 2007;3:138–153.
35. Heisteringer M, Stockenhuber F, Schneider B, Pabinger I, Brenner B, Wagner B, et al. Effect of conjugated estrogens on platelet function and prostacyclin generation in CRF. *Kidney Int* 1990;38:1181–1186.
36. Shemin D, Elnour M, Amarantes B, Abuelo JG, Chazan JA. Oral estrogens decrease bleeding time and improve clinical bleeding in patients with renal failure. *Am J Med* 1990;89:436–440.
37. Garay RP, Chiavaroli C, Hannaert P. Therapeutic Efficacy and Mechanism of Action of Ethamsylate, a Long-Standing Hemostatic Agent. *American Journal of Therapeutics*. 2006;13:236–247.
38. Hernandez MR, Alvarez-Guerra M, Escolar G, Chiavaroli C, Hannaert P, Garay RP. The hemostatic agent ethamsylate promotes platelet/leukocyte aggregate formation in a model of vascular injury. *Fundam Clin Pharmacol*. 2004;18:423–430.
39. Khawar H, Kelley W, Guzman N. Fresh Frozen Plasma (FFP) [Updated 2019 Jul 2]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2019 Jan-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK513347/>
40. Schulman S, Bijsterveld NR. Anticoagulants and their reversal. *Transfus Med Rev*. 2007;21:37–48.
41. Leissinger CA, Blatt PM, Hoots WK, Ewenstein B. Role of prothrombin complex concentrates in reversing warfarin anticoagulation: a review of the literature. *Am J Hematol*. 2008;83:137–143.
42. Franchini M, Lippi G. Prothrombin complex concentrates: an update. *Blood Transfus*. 2010;8:149–154.
43. Dutta TK, Verma SP. Rational Use of Recombinant Factor VIIa in Clinical Practice. *Indian J Hematol Blood Transfus*. 2014;30:85–90.
44. Hedner U. Mechanism of Action of Factor VIIa in the Treatment of Coagulopathies. *Seminars in Thrombosis and Hemostasis*. 2006;32:077–085.
45. Logan AC, Yank V, Stafford RS. Off-label use of recombinant factor VIIa in U.S. hospitals: analysis of hospital records. *Ann Intern Med*. 2011;154:516–522.
46. Tiede A, Worster A. Lessons from a systematic literature review of the effectiveness of recombinant factor VIIa in acquired haemophilia [published correction appears in *Ann Hematol*. 2018 ;97(12):2531]. *Ann Hematol*. 2018;97:1889–1901.
47. von Heymann C1, Jonas S, Spies C, Wernecke KD, Ziemer S, Janssen D, et al. Recombinant activated factor VIIa for the treatment of bleeding in major abdominal surgery including vascular and urological surgery: a review and meta-analysis of published data. *Crit Care*. 2008;12:R14. doi: 10.1186/cc6788. Epub 2008 Feb 15.
48. Stephan A Mayer, Nikolai C Brun, Kamilla Begtrup, Joseph Broderick, Stephen Davis, Michael N Diringer, et al. Efficacy and safety of recombinant activated factor VII for acute intracerebral hemorrhage. *N Engl J Med*. 2008;358:2127–2137.