GUIDELINES

Indian Antimicrobial Prescription Guidelines in Critically Ill Immunocompromised Patients


Keywords: immunocompromised patients, febrile neutropenia, solid organ transplant, Human Immunodeficiency Virus, Primary immunodeficiency disorders, asplenia


Source of support: Indian Society of Critical Care Medicine

Conflict of interest: None

INTRODUCTION

The number of admissions of immunocompromised patients in the Indian intensive care units (ICUs) is growing. This is because of availability of better treatments for acquired immunodeficiency states, increasing incidence and detection of cancer, with more aggressive therapies aimed at cancer cure, and increased expectations of better ICU outcomes in the cancer patient.1,2

The indications of immunosuppression have expanded, contributing to an increased number of immunocompromised patients. There is a sharp rise in the number of solid organ transplants being performed in India contributing to the increased number of patients with immunosuppression getting admitted to the ICU in immediate postoperative and subsequent follow-up period. In the pediatric population, we can recognize the genetic predisposition of patients to congenital immunodeficiency states. All these patients have greater susceptibility to new infections or reactivation of latent infections.3 Therefore, it is the need of the hour to develop Indian guidelines for prescribing antimicrobial therapy in this population presenting to the ICU.

We feel that the outcomes are likely to be better if these immunocompromised patients with infectious diseases requiring ICU admission are preferably treated at a tertiary care center where all diagnostic facilities along with specialists in microbiology, immunology, infectious disease are likely to be available. We present guidelines in five separate sections: febrile neutropenic patients, patients who have undergone a solid organ transplant, patients with human immunodeficiency virus infections presenting to ICU, congenital asplenia or hyposplenia, and those with congenital primary immunodeficiency syndromes.

These guidelines were developed starting with a collection of Indian and other data which was communicated to members by electronic communication. A meeting of all committee members was held where the data was presented in the form of questions pertaining to the evidence, available evidence from Indian ICUs, and evidence statements and recommendations. A draft of each section was prepared and circulated and finally presented in another committee meeting. The final draft of guidelines was prepared and again communicated to members. The GRADE system was used for the quality of evidence and recommendations (Table 1).

Table 1: Quality of evidence and recommendations

<table>
<thead>
<tr>
<th>Quality of Evidence</th>
<th>Level of evidence</th>
</tr>
</thead>
<tbody>
<tr>
<td>Evidence from ≥1 good quality and well conducted randomized control trial(s) or meta-analysis of RCT’s</td>
<td>1</td>
</tr>
<tr>
<td>Evidence from at least 1 RCT of moderate quality, or well-designed clinical trial without randomization; or from cohort or case-controlled studies.</td>
<td>2</td>
</tr>
<tr>
<td>Evidence from descriptive studies, or reports of expert committees, or opinion of respected authorities based on clinical experience.</td>
<td>3</td>
</tr>
<tr>
<td>Not backed by sufficient evidence; however, a consensus reached by the working group, based on clinical experience and expertise.</td>
<td>Useful Practice Points (UPP)</td>
</tr>
</tbody>
</table>

Corresponding author: Atul P Kulkarni, Professor and Head, Division of Critical Care Medicine, Department of Anaesthesiology, Critical Care and Pain, Tata Memorial Hospital, Homi Bhabha National Institute, Mumbai, Maharashtra, India. e-mail: kaivalyaak@yahoo.co.in
Indian Antimicrobial Prescription Guidelines in Critically Ill Immunocompromised Patients

PART 1. PATIENT WITH FEBRILE NEUTROPENIA IN THE INTENSIVE CARE UNIT

Febrile neutropenia (FN) is defined as an oral temperature of >38.3°C or two consecutive readings of >38.0°C for 2 hours and an absolute neutrophil count (ANC) of <0.5 × 10⁹/L or expected to fall below 0.5 × 10⁹/L.⁴

These guidelines are applicable to a critically ill febrile neutropenic patient presenting to the ICU, with any of the following clinical or laboratory parameters of organ failure but not limited to:

- Hypotension
- Tachypnoea requiring oxygen therapy more than 4 liters/minute to maintain saturation >90%
- Altered mental status (without focal neurological deficit)
- Oliguria or rising creatinine

Empiric Antibiotic Therapy in Critically Ill Febrile Neutropenic Patients with Suspected Bloodstream Infection

Evidence Statements

- In a critically ill febrile neutropenic patient presenting to the ICU with organ failure, empiric antibiotic therapy should be initiated or escalated to a broad spectrum carbapenem like imipenem or meropenem (UPP, A).
- Empiric combination of Imipenem or Meropenem and Colistin/Polymyxin B may be considered in the following patients having a high risk of infection with resistant gram-negative organisms (UPP, A):
  a. Critically ill patients with underlying acute Leukaemia presenting to the ICU.
  b. Patients of Leukaemia/Lymphoma on β-lactam/β-lactamase inhibitor (BL/BLI) ± aminoglycosides, shifted to ICU from the ward.
  c. Previous exposure to broad-spectrum antibiotics like carbapenem or BL/BLI combination in last 1 month.
  d. Hypotensive patients requiring vasopressor infusions (refractory septic shock).
  e. Patient shifted to the ICU on carbapenem therapy.
- We strongly caution against the use of an empiric combination of Imipenem or Meropenem and Colistin/Polymyxin B or Colistin/Polymyxin B alone in patients who are not a high risk of infection with carbapenem-resistant gram-negative organisms as defined above (UPP, A).
- We recommend against empirical use of Doripenem and Ertapenem (III, A)
- Vancomycin/Teicoplanin should be added as empiric therapy in a critically ill febrile neutropenic patient with
  a. Suspected Indwelling vascular catheter infection
  b. Skin and soft-tissue infection
  c. Previous Colonization/infection with methicillin-resistant Staphylococcus aureus (MRSA)
  d. Blood Culture: Gram-positive cocci awaiting identification
  e. Severe mucositis (III, A)
  f. Hemodynamic instability (hypotension) in patients admitted from home /OPD (UPP, A)
- We caution against empirical use of Vancomycin in another group of patients including patients on broad-spectrum antibiotics admitted to the ward and presenting to ICU with hemodynamic instability unless there is a high incidence of MRSA in the institute (UPP, A)

Evidence Summary

Common isolates in a blood culture in febrile neutropenic patients

In India, in febrile neutropenic patients, gram-negative bacteremia is much more common than gram-positive bacteremia (Table 2); in contrast to the western data, where gram-positive isolates are more common.⁵ ⁶ The spectrum of bacterial isolates from a number of studies in India suggest Enterobacteriaceae (E. coli and Klebsiella species) and Pseudomonas aeruginosa be the most common among gram-negative organisms. Among gram-positive isolates, Staphylococcus aureus and Coagulase-negative staphylococcus (CoNS) are most common isolates.

There are scarce data regarding the choice of empirical antibiotic regimens in critically ill febrile neutropenic presenting to the Indian ICUs. Most of the studies have a heterogeneous patient population–leukemia, lymphoma, solid, etc. Choice of antibiotics depends on the most likely causative microorganism, local antimicrobial sensitivity patterns, mechanism of action of antimicrobials, and site of infection, patient’s predisposing condition and treating physician’s judgment. Recent data show an increased prevalence of Multidrug-resistant (MDR) organisms. Several studies in India have shown that the majority of gram-negative bacteria isolated on initial blood cultures from patients were resistant to the non-carbapenem first-line antibiotics.⁴ ¹⁴ ¹⁷ Hence, initial antibiotic choice in a febrile neutropenic patient who is critically ill present-
ing to the ICU will be carbapenems like Meropenem or Imipenem. The prevalence of carbapenem-resistant gram-
negative organisms is alarming at present. According
to the Indian Council of Medical Research (ICMR) data
on non-neutropenic population, carbapenem resistance
among Enterobacteriaceae is 35 to 50 %, Pseudomonas spp
47% and Acinetobacter spp 62%. Based on the epide-
miology, current evidence and clinical experience the
committee has identified risk factors for carbapenem
resistance. Particular subgroups of patients, such as acute
leukemia patients presenting to the ICU, a patient already
on carbapenem shifted to ICU from the ward, previous
antibiotic exposure in the last 1 month and patients
on vasopressors are at risk of harboring carbapenem-
resistant organisms. Hence in these groups of patients,
initial empiric antibiotic regimen should include colis-
in/polymyxin B along with Imipenem or Meropenem
(Table 3).

Vancomycin is not a standard part of empirical anti-
biotic therapy for the febrile neutropenic patient. In the
western countries with predominant gram-positive bac-
teria and high incidence of MRSA, studies have failed
to show any benefit with empiric vancomycin in terms
of fever or mortality. In India, with predominant gram-
negative sepsis and low incidence of MRSA (35% as per
ICMR data), Vancomycin/Teicoplanin is recommended
as part of initial antibiotic regimen only in patients with
suspected indwelling catheter infection (rigors following
infusion, cellulitis at exit site), skin and soft tissue infec-
tion, severe mucositis, culture growing gram-positive
coci pending identification, previous MRSA coloniz-
/onction/ infection and hemodynamic instability admitted
from home/OPD.7

**Blood Culture Collection Procedure**

**Evidence Statements**

- We recommend a collection of at least two sets of
  blood cultures, with a set collected simultaneously
  from a peripheral site and one from the central line. In
the case of a multi-lumen catheter, one set per lumen
should be collected.
- Two blood culture sets from separate venepunctures
  should be sent if no central venous catheter is present.
Evidence Summary

The volume of blood is an important variable for detection of bloodstream infection volume of blood. Each ml of blood increased the yield (detection of positive culture) of blood cultures in adults by approximately 3%. Collection of two blood culture sets provide 80 to 90% yield of bloodstream pathogens in critically ill patients. In the pediatric population, smaller volumes of blood are suggested due to lesser total blood volume. The general consensus is not to exceed 1% of a patient’s total blood volume.

Empiric Antifungal Therapy in Febrile Neutropenic Patient

Evidence Statements

Following patients should be considered for initiation of antifungal therapy when they present to ICU with shock or respiratory distress especially when they have a persistent or recurrent fever or clinical deterioration after >3 days of broad-spectrum antibiotics (II, A)

- Allogenic hematopoietic stem cell transplantation (HSCT).
- Severe mucositis with diarrhea.
- Prolonged/anticipated duration of neutropenia >10 days.
- Worsening on broad-spectrum antibiotics like BLI and Carba-penems.
- More than 3 weeks of steroids.
- History of invasive fungal infection.
- New onset lung infiltrates (since chest X-ray has low sensitivity, high-resolution computed tomography (HRCT) should be done in these patients).

- We recommend the use of caspofungin (echinocandin group) as initial antifungal therapy. Caspofungin should be avoided in patients with chronic liver disease (Child-Pugh C).
- Anidulafungin and Micafungin can be considered if there are contraindications to use of caspofungin.
- Voriconazole is the drug of choice for proven, probable or possible aspergillosis. Due to its variable bioavailability, voriconazole should be administered IV. In patients with renal dysfunction, caspofungin can be given instead of IV voriconazole.
- Liposomal amphothericin B is the drug of choice for suspected or confirmed mucormycosis.
- All efforts should be made to confirm the presence of invasive fungal infection with the use of tests including CT chest, β-D-glucan, serum, and bronchoalveolar lavage (BAL) galactomannan, fungal culture. Tissue (lung/other clinically involved sites) biopsy should be performed if required, whenever feasible (II, A).
- We do not recommend the routine use of combination antifungal therapy for probable or proven invasive aspergillosis (III, A).

Evidence Summary

Invasive Candida or Aspergillus infections have been demonstrated in the autopsy of patients who died of a neutropenic fever with no clinical evidence of invasive fungal infection (IFI) except for a continuous fever. It is estimated that approximately 15 to 45% of patients with prolonged neutropenia have an invasive fungal infection (IFI). IFI is difficult to diagnose both in critically ill patients and patients with febrile neutropenia. Invasive fungal infection is associated with high mortality in both these groups especially if treatment is delayed.

High-risk patients who have received intensive cytotoxic chemotherapy are at risk for invasive fungal infection. Yeast (primarily Candida species) and molds typically cause infections, which are manifested by persistent or recurrent fever in patients with prolonged neutropenia, rather than causing initial fever in the course of neutropenia. Empirical antifungal therapy is instituted for the treatment of “occult” fungal infection presenting as persistent neutropenic fever despite 4 to 7 days of empirical antibiotic therapy.

The echinocandins have demonstrated significant fungicidal activity and treatment success against most of the Candida species in randomized clinical trials. Availability of intravenous formulation, limited drug interactions, favorable safety, and efficacy profile make them the first choice of empirical antifungal in critically ill patients including patients with febrile neutropenia. It is generally agreed upon that individual echinocandins namely caspofungin, micafungin and anidulafungin have similar efficacy and are interchangeable.

Recommended Dosage

- Caspofungin: loading dose of 70 mg followed by 50 mg daily. Dosage reduction is recommended for patients with moderate to severe hepatic dysfunction.
- Micafungin: 100 mg daily. No dose adjustment in the liver or renal failure.
- Anidulafungin: loading dose of 200 mg, followed by 100 mg daily. No dose adjustment is required in the liver or renal failure.

Limitations of echinocandin therapy should be kept in mind. Echinocandins have poor penetration in eye, CNS, and urine. Intuitively they should not be used for the treatment of fungal meningitis, endophthalmitis, and urinary tract infection. Echinocandins are not active against Zygomycosis.
**Echinocandins** have activity against *Aspergillus* species. However, **Echinocandins** monotherapy as the first line for treatment of *Aspergillus* is not studied well. **Echinocandins** have shown to be effective in salvage therapy however they are not recommended as monotherapy for the primary treatment of IA due to lack of evidence.29

Lipid formulations of amphotericin B should be used as first-line treatment if *Mucormycosis* (Zygomycosis) is suspected. The recommended dose is 3 to 5 mg/kg daily.32

Invasive aspergillosis should be suspected in patients with persistent febrile neutropenia with the development of signs of pneumonia including lung infiltrate. Voriconazole can be used for suspected or proven cases of IPA. The dose of Voriconazole is 400 mg (6 mg/kg) twice daily for two doses, then mg/kg) twice daily. As mentioned above, Echinocandins are recommended for salvage therapy of aspergillosis.33

**Amphotericin B** deoxycholate should be avoided in patients with underlying renal impairment, patients on other nephrotoxic drugs such as cyclosporine or tacrolimus after allogeneic HSCT, or antibiotics, such as aminoglycosides and in patients with previous history of toxicity.

Recommended minimum duration of therapy for candidemia without metastatic complications is 2 weeks after documented clearance of *Candida* from the bloodstream, provided neutropenia and symptoms attributable to candidemia have resolved.7

Investigation for confirmation of invasive fungal infection:

- Poor sensitivity of chest radiograph compared to CT scan for detection of pneumonia in this population should be kept in mind.34 The galactomannan assay is highly specific for *Aspergillus* species with some cross-reactivity with *Histoplasma capsulatum* and *Penicillium* species. The false positive reaction can occur with concomitant use of β-lactam/β-lactamase combinations, such as piperacillin-tazobactam.22

Marr et al have demonstrated 8.2% absolute reduction in mortality rates with the combination of voriconazole and anidulafungin in adult patients with hematologic malignancies (HMs) and hematopoietic stem cell transplantation (HSCT) having probable or proven Invasive aspergillosis (IA). However, this difference was not statistically significant.

Combination antifungal treatments are used with the rationale to maximize treatment by targeting multiple sites or metabolic pathways or different steps in the same pathway hence leading to an additive or synergistic effect. While in vitro studies on combination antifungals showed additive or synergistic effect; in vivo studies have given mixed results. While the combinations of azoles with echinocandins has shown additive or synergistic effects; combinations of polyenes with triazoles have demonstrated the antagonistic effect. Lack of in vitro findings has not been replicated with consistency. These in vivo studies are difficult to interpret due to the lack of validated protocols, variable test designs, and a definite end point. Combination antifungal therapy definitely increases the cost of treatment, has the potential to cause deleterious drug interactions, may add toxic effects and have the possibility of increased risk for resistance. In view of the lack of strong evidence in favor as well as potential harmful effects, routine use of a combination of combination antifungal therapy is not recommended.35

### Empiric Antiviral Therapy in Febrile Neutropenic Patient

**Evidence Statements**

- There is no role of empirical antiviral therapy with febrile neutropenia (III, A)
- Acyclovir is recommended for the treatment of Varicella-Zoster virus (VZV) or Herpes Simplex virus (HSV) infection in the presence of clinical or laboratory evidence of infection is present (III, B)
- Serological (Immunoglobulin) tests should not be used to diagnose VZV or HSV infection (III, A)
- Ganciclovir is recommended for the empiric therapy for Cytomegalovirus (CMV) in below-mentioned patients with high risk of CMV reactivation if they present with diarrhea/pneumonia not responding to antibiotics and antifungals (UPP , A)
  - a. Patients on T-cell-depleting agents such as fludarabine/purine analogs as well as rituximab
  - b. Patients on high dose steroids who develop diarrhea
  - c. Pneumonia not responding to antibiotics and antifungals

**Evidence Summary**

General consensus does not recommend the empirical use of antiviral drugs in the management of febrile neutropenic patients except in cases with clinical or laboratory evidence of viral infection. Cellular immunity offered by T cells helps in controlling CMV replication preventing its recurrence hence patients receiving T-cell-depleting agents are at risk of CMV reactivation. No specific treatment for infections with RSV and parainfluenza viruses is recommended due to the lack of specific evidence.40,41
Empiric Treatment Against Pneumocystis Carinii/Jiroveci Pneumonia (PCP) in febrile neutropenic patient

Evidence Statements
- Treatment with sulfamethoxazole/trimethoprim should be considered in high-risk patients such as allogeneic HSCT, high-dose corticosteroid therapy administration of T-cell-depleting agents such as fludarabine/purine analogs and rituximab when such patients present with hypoxemic respiratory failure with or without radiological evidence of pneumocystis carinii pneumonia especially if they are not on PCP prophylaxis (III, A)
- Every attempt should be made to confirm PCP infection (I, A)

Evidence Summary
Patients considered high risk for PCP infection are allogeneic HSCT recipients (Good evidence), autologous HSCT, high-dose corticosteroid therapy and patients receiving T-cell-depleting agents such as fludarabine, purine analogs and rituximab (moderate evidence).42-44 If they present with hypoxemic respiratory failure, then such patients should be on sulfamethoxazole/trimethoprim. Hypoxemia is the most characteristic abnormality in PCP pneumonia. Chest radiograph might be normal in early disease.

Role for Empiric Antimicrobial Therapy for Tropical Infections like Malaria, Leptospirosis

Evidence Statements
- There is no role for empirical antimicrobial therapy against tropical infections like malaria, leptospirosis (III, A)
- Documented tropical infections in neutropenic patients in ICU should be treated similarly as they are treated in non-neutropenic patients (UPP, B)

Evidence Summary
There are occasional reports of malaria in patients on chemotherapy with a solid tumor and hematolymphoid malignancies with febrile neutropenia. In a series of 99 patients of acute leukemia on chemotherapy with febrile neutropenia, malaria was responsible for fever in only 4% of patients.45

Febrile neutropenia patient presenting to ICU often have thrombocytopenia due to the disease itself, chemotherapy or sepsis. Presence of fever and thrombocytopenia itself should not warrant empirical anti-malarial therapy even in a malaria-endemic country like India. A high index of suspicion is warranted in a resident or traveler of malaria endemic area who presents with the classic triad of symptoms (fever, chills, and sweating). If malaria is suspected, peripheral smear for malaria parasite and rapid malaria antigen [e.g. Histidine-rich protein II (HRP-II) antigen of Plasmodium falciparum and common Plasmodium lactate dehydrogenase (pLDH) of Plasmodium species] should be performed early, and antimalarial therapy should be initiated in positive cases. With rapidity (diagnosis in less than an hour) and good negative predictive value of (98.2%) malaria antigen test, antimalarial therapy is restricted only to positive cases.46

There is lack of enough evidence documenting the etiological role of other tropical infections like Leptospirosis in a subset of patients with febrile neutropenia; hence we believe that until enough evidence is available, suspected or documented tropical infections in neutropenic patients in ICU should be treated similarly as they are treated in non-neutropenic patients.

Role of Surveillance Cultures

Evidence Statement
- We strongly recommend against repeated surveillance cultures as these do not help to guide antibiotic therapy (III, A)

Evidence Summary
As most of the infections in neutropenic patients occur due to organisms in the respiratory or gastrointestinal tract, therefore surveillance culture seems to be a reasonable strategy in deciding the empiric antibiotic therapy in febrile neutropenia. The studies published in the 1980s and 90s supported the practice of surveillance culture. However, there has been a very poor correlation between blood and fecal isolates in most of the studies.47,48

Widespread antimicrobial treatment may inhibit the growth or distort the proportion of different species found in fecal cultures. A recent study conducted in pediatric allogeneic HSCT patients has demonstrated a positive predictive value of 0.9% to bacterial surveillance cultures, with a sensitivity of 33.3% and a specificity of 47.4%. Surveillance cultures were not cost effective. The sampling and analyses require lots of laboratory and nursing resources.49 Another study in adults who got admitted for HSCT concluded that surveillance blood cultures in patients who have undergone HSCT do not identify bloodstream infections. The number of positive blood cultures was not helpful in determining which patients had an infection.50
Role of Source Control in the Treatment of a Febrile Neutropenic Patient

Evidence Statements

- We recommend that in patients with febrile neutropenia with a clinically documented source of infection; immediate intervention should be undertaken for source control (III, A)
- We recommend that Non-tunnelled (short term) central venous catheter (CVC) should be promptly removed in following cases (III, A).
  a. CVC is an obvious source of infection.
  b. Patient not improving/deteriorating in spite of 24 hours of antibiotics and resuscitation with no other source of infection evident
- We recommend that long term central venous catheter should be promptly removed in presence of any of the following (III, A).
  a. Documented line/blood infection with S. aureus, or Candida species
  b. Tunnel infection or port abscess
  c. Septic thrombosis, endocarditis or osteomyelitis
- We suggest considering the removal of long term central venous catheter in the absence of above-mentioned features if there is deterioration or no improvement in patients condition and no other obvious source of infection is evident (UPP, B)

Evidence Summary

Control of source in the form of drainage of an abscess, debridement of infected necrotic tissue and removal of a potentially infected device is of paramount importance. Foci of infection readily amenable to source control include but not limited to intra-abdominal abscesses, gastrointestinal perforation, ischaemic bowel or volvulus, cholangitis, cholecystitis, pyelonephritis associated with obstruction or abscess, necrotizing soft tissue infection, empyema, septic arthritis, and implanted device infections. There is general agreement that source control should be done at the earliest to reduce microbiological burden, and mere antibiotics and resuscitation would not achieve cure unless adequate source control is done. If Vascular catheters are suspected, its prompt removal should be considered. It is important to note that the classical clinical signs of infection (ruber, calor, dolor, etc) be absent due to low neutrophil counts.

Patients with S. aureus catheter-related bloodstream infection (CRBSI) have a significantly higher risk of hematogenous complications with the retained foreign body, especially in immunocompromised patients. Retention of the catheter has shown to worsen the outcome of candidaemia.

Antibiotics De-escalation

Recommendations

- Antimicrobial de-escalation should be implemented in the following situations: (III, A)
  a. Once and if a pathogen is identified, we recommend de-escalation to an antibiotic that the organism is susceptible to.
  b. We recommend treating with appropriate agents based on the site and pathogen until the patient is afebrile for at least 48 hours and there is evidence of marrow recovery (neutrophil count ≥ 500 cells/mm³).
  c. In patients without microbiologically documented infection we recommend continuing empirical antimicrobials until the patient is afebrile for at least 48 hours and there is evidence of marrow recovery (neutrophil count ≥ 500 cells/mm³)

Evidence Summary

Data on de-escalation strategies in neutropenic patients after identification of a clinically relevant pathogen is scant, and there is no data on de-escalation when no pathogen has been identified. Although antibiotics are required to treat an occult infection during neutropenia, marrow recovery is necessary to protect the patient.

Antibiotics for Multidrug-resistant Bacteria

Useful Practice Points

Antibiotics like Fosfomycin, tigecycline and minocycline may be considered in infection with multidrug-resistant bacteria in the presence of in vitro susceptibility after considering the in vivo penetration at the source of sepsis.

<table>
<thead>
<tr>
<th>Good activity</th>
<th>Bad activity</th>
</tr>
</thead>
<tbody>
<tr>
<td>E. coli, Citrobacter sp, Proteus mirabilis, Streptococcus sp, Staphylococcus aureus (MSSA and MRSA) and Enterococcus faecalis</td>
<td>Pseudomonas sp, Acinetobacter sp, Bacteroides (Anaerobes), Coagulase negative staphylococcus, Morganella- morganii and Enterococcus faecium</td>
</tr>
</tbody>
</table>

Evidence Summary

- Most Enterobacteriaceae, Acinetobacter sp, Staphylococcus aureus (MSSA, MRSA), Enterococcus sp (VRE) and anaerobes
- Fosfomycin
- Tigecycline
- Minocycline
- Most Enterobacteriaceae, Acinetobacter sp, Staphylococcus aureus (MSSA, MRSA), Enterooccus sp (VRE) and anaerobes

S70
**Indian Antimicrobial Prescription Guidelines in Critically Ill Immunocompromised Patients**

<table>
<thead>
<tr>
<th>Uses</th>
<th>Bacteraemia, VAP, UTI, wound infection and meningitis</th>
<th>Skin and soft tissue infection and intra-abdominal infection</th>
<th>Ventilator-associated pneumonia (VAP)</th>
<th>Urinary tract infections (UTI)</th>
<th>Hospital/healthcare-Associated Pneumonia (HAP)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Caution</td>
<td>High sodium load</td>
<td>Should not be used as a sole agent in Bacteremia, pneumonia</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**REFERENCES**


2. SOLID ORGAN TRANSPLANT (SOT) RECIPIENT IN THE INTENSIVE CARE UNIT (ICU)

Infectious complications in solid organ transplant (SOT) recipients pose unique challenges when such patients require intensive care unit (ICU) care. The net immunological state of these patients is a measure of an individual’s unique susceptibility to infection and incorporates an assessment of several important contributing factors.

- Pretransplant diagnosis or treatment
- Nature of organ transplant received (e.g., lung vs. liver organ transplant)
- Dose, duration, and choice of maintenance immunosuppression
- Comorbidities (e.g., viral coinfection [hepatitis C virus (HCV), cytomegalovirus (CMV)], malnutrition, end-organ failure [cirrhosis, chronic kidney disease])
- Breaches of the mucocutaneous barrier: Indwelling devices, mucositis

Detection of infection in SOT recipients is difficult and requires a high index of suspicion, as fever and localizing signs are usually absent in the majority of the patients. Detection of infections should include an assessment of risk factors, detailed history, and physical examination. The infections in SOT patients can be categorized as follows.

- C–Community-acquired
- R–Reactivation
- E–Epidemiologic exposure
- D–Donor-derived
- I–Iatrogenic
- T–Travel related

It is advisable to have a syndrome-based approach (e.g., Nonspecific febrile illness, pneumonia, urinary tract, central nervous system) at first and then narrow the differential diagnoses of possible organisms that could cause the clinical presentation.

Microbiological diagnosis is crucial in this patient group. In the context of extensive differential diagnoses, the value of early and specific diagnostics with the use of invasive procedures if necessary (bronchoscopy, tissue biopsy, or aspiration of collections) to obtain specimens cannot be underestimated. After transplantation, serologic techniques are of limited use because transplant recipients may not mount timely serologic responses.

### Table 1: Timeline of infections after a solid organ transplant

<table>
<thead>
<tr>
<th>Infection characteristics</th>
<th>Timeline</th>
<th>Infection Types</th>
<th>Usually community-acquired infection</th>
<th>Reactivation of latent infections during immunosuppression</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0-1-month Post-SOT</td>
<td>Nosocomial infection</td>
<td>Opportunistic Infections,</td>
<td>Reactivation of latent infections</td>
</tr>
<tr>
<td></td>
<td>1 to 6-months Post-SOT</td>
<td>Pneumonia-HAP, VAP, CRBSI, CAUTI, Post-surgical Infections, Donor-derived infections</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>More than 6-months Post-SOT</td>
<td>M. tuberculosis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bacterial</td>
<td>Resistant Bacteria</td>
<td>MRSA, VRE, MDR gram-negative bacilli</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>C. difficile associated infections</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Viral</td>
<td>HSV (in absence of anti-HSV prophylaxis)</td>
<td>BK Virus, Adenovirus</td>
<td>Recurrent HSV, VZV</td>
<td></td>
</tr>
<tr>
<td></td>
<td>HIV</td>
<td>RSV</td>
<td>Adenovirus</td>
<td></td>
</tr>
<tr>
<td></td>
<td>West Nile virus</td>
<td>HCV Reactivation CMV, EBV, HSV, VZV</td>
<td>HCV Reactivation</td>
<td></td>
</tr>
<tr>
<td>Fungal</td>
<td>Candida (likely to be Fluconazole resistant)</td>
<td>Aspergillus Cryptococcus Zygomyces</td>
<td>During intense immunosuppression: Aspergillus Cryptococcus Zygomyces PCP</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Early Aspergilosis (Uncommon)</td>
<td>PCP if not on prophylaxis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Parasitic</td>
<td>Rare</td>
<td>Toxoplasma Strongyloides Leishmania, Trypanosoma</td>
<td>Ongoing risk of Toxoplasma Strongyloides Leishmania, Trypanosoma if immunosuppression intensified</td>
<td></td>
</tr>
</tbody>
</table>
Thus, antigen detection or molecular nucleic acid detection assays are preferred over serologic testing.

The timeline of post-transplant infections reflects the post-transplantation relationship between the recipient’s epidemiologic exposures and immunosuppressive strategy employed (Table 1). It is used to establish a differential diagnosis for infectious syndromes at various stages after transplantation. Infections occurring outside the usual period or of unusual severity suggest excessive immunosuppression or epidemiologic hazard. Most centers use a variation of standard ‘triple immunosuppression’ (prednisone, calcineurin inhibitor, antimetabolite such as mycophenolate mofetil).

Evidence Statements

• Infections in the first month (0–30 days) of post SOT period should be treated similarly to the treatment of a non-immunocompromised postoperative patient (I, A).

• Approach to initial diagnostic workup and empiric therapy in post SOT recipients (30–180 days).

• We recommend early BAL in patients with suspected pneumonia coming to ICU (I, A). We recommend that the BAL fluid should be tested for (I, A).

• Total and differential cell counts

Microbiology

- **Cultures**: Aerobic culture for bacteria, mycobacterial growth indicator tube (MGIT) for mycobacterium tuberculosis, fungal culture
- **Stains and immunohistochemistry**
  - Gram stain: Bacterial
  - KOH preparation/Calcofluor white: Fungal
  - Auramine-rhodamine, auramine-o, or ziehl-neelson:

- **Modifed acid-fast stain (kinyoun): Nocardia**
- **Silver methenamine: Pneumocystis carinii pneumonia, fungal**
- **Galactomannan assay**: Negative predictive value <0.5, positive predictive value >3
- **Polymerase chain reaction (PCR)**
  - *Mycobacterium tuberculosis*: Cartridge Based Nucleic Acid Amplification Test (CB-NAAT or GeneXpert)
  - Multiplex PCR assay
- **Quantitative or semiquantitative cultures particularly for pneumonia.**

Following organisms are diagnostic of infections. If organisms are identified it is less likely to be the contaminants/colonizers and should be treated.

- **Pneumocystis carinii**
- **Toxoplasma gondii**
- **Strongyloides stercoralis**
- **Legionella pneumophila**
- **Cryptococcus neoformans**
- **Histoplasma capsulatum**
- **Mycobacterium tuberculosis**
- **Mycoplasma pneumoniae**
- **Influenza a and b viruses**
- **Respiratory syncytial virus**
- **Cytology**

• We recommend HRCT chest for the diagnosis of nodular infiltrates to rule out invasive aspergillosis (I, A).

• We recommend empiric management with carbapenem and azithromycin till causative organisms are identified (I, A).

• We recommend adding polymyxins if the patient is admitted with ≥ 48 hours before symptoms (I, A)

Medical care (hemodialysis, wound care, immunosuppressants) within the previous 30 days (UPP, B)

• Hospitalization in an acute care hospital ≥ 2 days within the prior 90 days (UPP, B)

• We recommend voriconazole in the dose of 6 mg/kg bd for 1-day t/b 3mg/kg bd (I, A)

• As a salvage therapy, we recommend Caspofungin (dose 70 mg IV followed by 50 mg iv daily) (I, A)

• We recommend therapeutic drug level monitoring of the voriconazole when using these agents for the treatment of Invasive Aspergillosis (I, A).

Evidence Summary

During the first month after SOT, opportunistic infections are generally absent as the full effect of immunosuppression has not yet been established. In this period donor-derived or recipient-derived viremia, candidemia or technical complications related to surgery are common. Most infections are caused by gram-negative bacterial (GNB). Bacterial infections are either nosocomial or healthcare-associated (27.4% and 49.8% respectively); the remaining 22.9% are community-acquired. The urinary tract infection (UTI) is the most common primary source of GNB infections in 55.2% of SOT recipients. Gastrointestinal infections are the second most common infections (15.2%). Other infections include infections of the respiratory tract (3.6%), intravascular catheters (3.6%) and skin and soft tissue (2.7%) are also seen in SOT recipients in decreasing order of occurrence. E. coli has been found to be the most common GNB (36.8%), followed by *K. pneumoniae* (14.3%), *P. aeruginosa* (13.0%), *Enterobacter cloacae* (4.9%) and *Citrobacter freundii* (4.5%) whereas polymicrobial infections occurred in 8.1% of cases. The incidence of GNB infections shows a declining trend to fall to 25.7% (95% CI: 20.1–32.1) from 2 to 12 months. It further declines to 8.2% (95% CI: 6.7–10.0) after 12 months per 1000 person-years following SOT.1,2
One prospective multicenter study involving 35 centers reported the incidence of pneumonia after SOT as 10.1 episodes/1000 recipients/year and bacteria (87.1%), virus (29%), and fungi (6.4%) were the common causative agents. A multidrug-resistant bacterium is isolated in 18.2%, 40%, and 100% of patients with CAP, HCAP, and HAP (p = 0.007), respectively. Overall, 11.1% of patients admitted to the intensive care unit, 3.7% developed graft rejection, and graft function deteriorated in 18.5%. In-hospital mortality was 1.9%. In renal transplant recipients, bacterial infections are the most common cause of pneumonia. A retrospective study of 406 renal transplant recipients observed that incidence of healthcare-associated pneumonia is 56% and bacterial infections are the most common cause (31% of the patients), especially *haemophilus influenzae*, *staphylococcus aureus*, *gram-negative bacilli* (33.3%), and *pseudomonas aeruginosa*. Another prospective study of 610 kidney transplant recipients also observed 60 episodes of pneumonia in 54 patients (8.8%), of which 23 had a nosocomial origin (38%) and 37 were community-acquired infections (62%). Bacterial infection is the most frequent etiology (44%), followed by fungal in 4 (7%) and viral in 2 (3.5%). The most commonly isolated microorganism in nosocomial pneumonia is *P. aeruginosa* (26%, among which 50% were multidrug resistant). In 34% episodes, no microorganism is isolated. The most common pathogen among community-acquired pneumonias is *S. pneumoniae* (11%). In 54% of cases, there is no microbiologic confirmation of disease. The overall accuracy of bronchoalveolar lavage is 72%. When 21 patients with pneumonia (35%) were admitted to the ICU; of these, 14 had a nosocomial infection (60%) and 9 (15%) died due to the infection (p = 0.001).

Data on heart transplant recipients has also shown that pneumonia is the most common complication. A prospective review of 307 heart transplant recipients found 21.1 cases of pneumonia per 100 heart transplantations. 75% of the cases occurred in the first 3 months, 82 causal agents are identified, of which 60% were opportunistic,
25% are nosocomial, and 15% were community-acquired. The most frequent isolates were CMV,20 aspergillus species,13 and pneumocystis carinii.11 Hemoptysis occurred more frequently in aspergillus pneumonias than in other pneumonias (54% vs. 6%, respectively; p < .05); aspergillus pneumonia is the only type of pneumonia during which cavitated nodules were noted on thoracic radiographs. The overall mortality rate was found to be 30.8%.10 Lenner et al reported 47 of 159 heart transplant recipients (29.9%) had 81 pulmonary complications. 11Including Pneumonia (n = 27), and bronchitis (n = 15).11

A retrospective review of 34 heart transplant recipients (31.3%) who developed pulmonary complications, within first 6 months post-transplant showed Hospital-acquired bacterial pneumonia in 5 patients, fungal pneumonia in 3 patients, a post-transplant lymphoproliferative disease in 1 patient, and community-acquired pneumonia in 1 patient. Pneumonia-related mortality rate was 14.7% due to early-onset nosocomial pneumonias where bacterial and opportunistic microorganisms organisms were more commonly seen.12

SOT recipients are at risk of developing bacterial infections like nocardia.13 The risk of developing nocardiosis after SOT varies with the type of organ transplanted, e.g. lung transplant. In a review of 5126 organ transplant recipients has demonstrated that highest nocardial infection rate was seen among lung transplant recipients (3.5%).14

Due to frequent exposure to antibiotics and repeated hospitalizations; SOT patients are at risk of developing intra-abdominal infections (IAI). IAI is commonly seen as a complication in post-liver, pancreas, multi-visceral transplants. Superinfection with MDR pathogens occurs frequently causing tertiary peritonitis.15 Clostridium difficile-associated disease (CDAD) is the most common cause of nosocomial diarrhea. One cohort study involving 4472 SOT patients observed that 42 episodes of CDAD were diagnosed in 36 patients (0.94%). Median onset of infection was 31.5 days (range 6–741). It occurred during the first month after transplantation in half the cases and overall the prognosis was good.16

SOT recipients are at risk to develop viral infections leading to various nonspecific viral syndromes. Common viruses seen are CMV, EBV, and other viruses like herpesvirus,6 Zika virus, RNA respiratory viruses, adenovirus, norovirus, and polyomaviruses.17 Among viruses CMV virus disease which occurs during the first 3 months. With the introduction of CMV prophylaxis, this onset has been delayed. The seroprevalence rate of CMV ranges from 30–97%.18

The incidence of PCP in SOT recipients is variable. In a retrospective study of 1192 renal transplant patients, it was reported to be 0.6 to 9%. They observed that the incidence of PCP with a moderate Cyclosporine based immunosuppressive regimen is low and seems to occur only in cases of additional immunosuppressive cofactors.19
In another retrospective study of 601 renal transplant recipients, PCP incidence was 2.2%.20 In liver transplant recipients (154 adult patients) PCP occurred in 5.2% and the authors observed that patients who developed PCP had more episodes of rejection (p < 0.05), received more OKT3 (p < 0.05), a prednisone (p < 0.05) than controls.21

Another retrospective study of 43 adult OLT recipients showed that the incidence of PCP was 0.9%. Most of the patients developed PCP at around 1 year of post-OLT, and the risk of PCP was closely related to strong immunosuppressive regimen. Thus they advised that routine PCP prophylaxis for 12 months be continued for 12 months, among patients receiving antirejection treatment.22

Invasive fungal infection (IFIs) frequently complicates post SOT course. Cryptococcosis is a significant opportunistic infection in SOT recipients following aspergillosis and candidiasis. CSF analysis is highly recommended to diagnose underlying CNS disease in suspected cases.23
In Lung and heart transplant, ~ 2% incidence IFI of lung was noted and even dissemination can occur. Cryptococcus can colonize the airways of lung transplant recipients and can cause endobronchial fungal infection. It can present with skin manifestations and Immune reconstitution syndrome as well.24,25 In heart transplant recipients incidence of cryptococcus is 3%, it manifests as sepsis and is associated with high mortality.26,27 In renal transplant recipients incidence of cryptococcus is 2.8% manifests as cryptococcal necrotizing soft tissue infection.28 In liver transplant recipients the incidence is 2.4% and liver failure is independently associated with Cryptococcal meningitis mortality.29 However, with wider employment of antifungal prophylaxis and improvements in transplantation practices, there is a decline in the overall incidence of IFIs.30,31 SOT recipients have risk of developing aspergillus particularity in lung transplant. A retrospective study involving 362 lung transplant recipients found that 105/335 (31%) patients had evidence of aspergillus infection (colonisation or invasion), 83 (25%) patients had colonisation and 22 (6%) patients had radiographic or histological evidence of invasive disease. Most of the infections occurred within the first 3 months after transplantation. Invasive aspergillosis (IA) was associated with 58% mortality after 2 years, while colonisation was associated with increased mortality after 5 years compared non-colonised patients (p < 0.05).32 Hambrecht et al compared voriconazole with amphotericin b in their large randomized trial for the treatment of invasive aspergillosis (IA) in immunocompromised patients.33
After this trial voriconazole had been considered the preferred agent for IA. In their study they found that at week 12, there were more successful outcomes 52.8% patients in the voriconazole group (complete response 20.8% and partial response in 31.9%) compared to 31.6% in the amphotericin B group (complete response 16.5% partial response in 15%). The survival rate was better at 12 weeks in voriconazole compared to amphotericin B group. (71% vs. 58%) (HR- 0.59; 95% CI- 0.40 to 0.88).

After this study several studies were conducted for use of voriconazole for the treatment of IA, especially in patients of solid organ transplant recipient. J. Fortun et al in their case series reported of 100 % complete response in four patients (two liver transplant, one lung transplant and one kidney transplant) of IA after treatment with voriconazole. Denning et al in their study showed good response in IA treated with voriconazole; 56 out of 60 patients in voriconazole group were treated successfully. Veroux et al reported complete response in four kidney transplant patients with IA treated with voriconazole. In another report that included 11 SOT recipients with central nervous system aspergillosis treated with voriconazole, the favorable response rate was 36% and survival was 31%. Voriconazole was successfully used in heart transplant recipients as first-line and salvage therapy for IA. Plasma drug level monitoring is important when voriconazole is used as the plasma levels achieved are variable and often do not reach therapeutic levels in the plasma, requiring dose adjustments. The fact that clinical efficacy is dependent on the achievement of therapeutic drug levels has been well established.

CMV management

- We recommend IV ganciclovir 5mg/kg twice a day as the initial treatment for (I, A):
  a. Severe or life-threatening CMV disease
  b. Patients with high and increasing viral load
  c. Patients with questionable gastrointestinal absorption
- Oral valganciclovir 900 mg once a day is an effective initial therapy for mild to moderate CMV disease (I, A), or as a step down to iv ganciclovir after clinical improvement (II, B)
- We recommend against the use of acyclovir and oral ganciclovir for treating CMV disease (UPP, A).
- IV gamma globulin or preferably CMV-specific gamma globulin if available in the dose of 1 gm/kg over two days may be considered for patients with life-threatening disease, CMV pneumonitis (II, B).
- We recommend a duration of treatment for a minimum of two weeks. It should be continued until viral eradication is achieved either by weekly monitoring for viral load by real-time quantitative PCR (I, A).
- Therapy should be extended beyond two weeks if the clinical resolution is not seen or virus load continues to be high (I, A).
- After completion of full-dose antiviral treatment, a 1 to 3 months course of secondary prophylaxis may be considered depending on the clinical situation (II, B).
- Cautious reduction in immunosuppression should be considered in SOT patients presenting with CMV disease, especially if the disease is moderate to severe (II, B).

Evidence Summary

Drugs used for the treatment of CMV disease are IV Ganciclovir and oral valganciclovir. Oral ganciclovir should not be used due to poor bioavailability. Ganciclovir through IV route has been demonstrated as the treatment of choice in many trials. Asberg et al in a randomized controlled trial compared the outcome of CMV disease after treatment with IV Ganciclovir and oral valganclovir. Three hundred twenty-one SOT recipients were enrolled and randomized to receive either twice daily intravenous ganciclovir or oral valganclovir for 21 days followed by once daily valganclovir until day 49 in all the patients. All patients were followed up for 1 year. The success rate was the same in both the groups with a similar rate of clinical and viral eradication. The clinical recurrence rate was also not statistically different in both the groups. In a retrospective study, the response to therapy was assessed using RT-PCR (2262 samples) and antigenemia using pp65 assay (1285 specimens). Both methods had > 90% specificity, but RT-PCR had better sensitivity. The authors concluded that RT-PCR was a more reliable tool to monitor the response to therapy. Failure to eradicate DNA-emia was the only independent predictor of recurrence in both the groups. The efficacy for the eradication of viremia was similar in both the groups, still many patients were viraemic even after treatment for 21 days, hence, duration of treatment should be individualized and based on clinical resolution and virologic clearance. There is a direct association between viral suppression below the lower limit of quantified test and disease resolution. Rapid resolution of CMV disease is seen with lower pre-treatment viral load (lower than 18,200 IU/mL).

PCP Management

- We recommend trimethoprim-sulfamethoxazole (TMP-SMX) as the first-line agent and drug of choice with the Trimethoprim component being 20 mg/kg /day in 3 to 4 divided doses (I, A).
- In severe infections, if available, intravenous pentamidine probably remains the second-line agent after TMP-SMX (II, A).
• In patients with hypoxemia (PaO₂ < 70 mmHg on room air), adjunctive corticosteroids should be administered with antimicrobial therapy, ideally within 72 hours of initiating antimicrobial therapy for maximum benefit (II, A). The dose of steroids should be 1 mg/kg/day prednisone (or equivalent) given in two divided doses daily for 5 to 7 days (II, A). Steroids should be tapered over a period of 7 to 14 days (II, B).
• Duration of antimicrobial therapy should be for at least 14 days (I, B).

Evidence Summary
TMP-SMX acts by interfering with folate metabolism and remains the drug of choice for treatment of PCP in SOT patients, HIV patients, and non-HIV patients. TMP-SMX has high efficacy and availability in both oral and IV preparation with good oral bioavailability too. Intravenous pentamidine has been found to be equally effective in HIV-infected patients and remains the second line of choice for treatment of PCP in SOT patients. However, the use of pentamidine has been largely limited in view of its numerous toxicities in 71% patients leading to withdrawal in around 18% patients. The optimal duration of therapy is usually 14 days which can be extended to 21 days in severe cases with slow clinical improvement. Adjunctive glucocorticoids are recommended for HIV-positive patients with moderate to severe PCP, defined as PaO₂ < 70 mmHg while breathing ambient room air. The benefit in survival from corticosteroids begins during the first 72 hours of treatment. Bolée et al’s study found a trend for longer survival in patients who received adjunctive steroids (p = 0.07). In Pareja et al’s study, there was no difference in the mortality rates of patients treated with adjunctive high-dose steroids, but they did spend less time on mechanical ventilation compared to patients not managed with steroids.

Tuberculosis in SOT Recipient

Evidence Statements
• The diagnosis of active TB in transplant recipients requires a high index of suspicion. Although the diagnostic modalities and treatment of TB in SOT patients remains the same as that in immunocompetent hosts, these individuals often require an invasive procedure, such as bronchoscopy with bronchoalveolar lavage or lung biopsy (I, A).
• Rifamycins, particularly rifampin, reduce serum concentrations of tacrolimus, cyclosporine, rapamycin (sirolimus), and everolimus via induction of the cytochrome p450 isoenzyme CYP3A4, necessary dose adjustments, and therapeutic drug monitoring are warranted to avoid development of rejection (II, A). When rifampin is not used, a longer than usual duration of treatment is required (II, B).

Evidence Summary
Given that tuberculosis is an immunological disease and with the high prevalence of TB in India, the incidence of active tuberculosis infection is higher among SOT recipients as compared to the general population. The diagnosis of TB in SOT recipients presents challenges that may lead to treatment delay. These include atypical clinical presentations, increased likelihood of negative tuberculin skin tests and/or IGRA, and negative sputum smear results despite active disease makes TB diagnosis in SOT recipients a challenge. One-third to one-half of cases of tuberculosis after transplant are disseminated or extrapulmonary. Lung transplant recipients are most likely to develop pulmonary manifestations of TB. Similarly, drug-drug interactions between immunosuppressive and AKT, allograft-related drug toxicities, and inadequate immune responses to TB makes treatment of TB in transplant recipients also very challenging.

Infective diarrhea in SOT patients

Evidence Statements
• We recommend empiric management of GI infections with ceftriaxone iv + ganciclovir 5g/kg BD IV and vancomycin 125 mg PO QID (if the patient is already on antibiotics to cover CDI) till definitive diagnosis is made (I, A).
• If the patient is in septic shock, based on local resistance pattern, and previous drug history of patient consider carbapenems (UPP, A).
• We recommend cessation of the inciting antimicrobial agent whenever possible (II, A).
• We recommend NAAT for the diagnosis of CDI (I, A).
• For mild-to-moderate CDI we recommend oral metronidazole (I, A). Dose of metronidazole 500 mg TID for adults.
• We recommend oral vancomycin for the treatment of severe CDI (IA). The accepted dose of vancomycin is 125 mg qid for adults and 40 to 50 mg/kg/day divided QID for pediatric patients (not to exceed adult dosing).
• In severe CDI with complications, a dose of oral vancomycin may be increased up to 500 mg orally QID.
• Vancomycin may be administered by retention enema (IIB), and intravenous metronidazole may be added (IIC).
• Feecal transplant may be considered in recurrent or relapsing CDI (II, B).
We suggest consideration for surgical intervention in cases of complicated CDI (II, B).
In cases of multiple recurrences of CDI, we recommend prolonged courses of oral vancomycin, either in a tapering or pulse dose schedule (II, A). Fidaxomicin can be used if available (II, B).

**Table 3: Common causes of diarrhea in SOT patients**

<table>
<thead>
<tr>
<th>Common</th>
<th>Unusual</th>
<th>Rare</th>
</tr>
</thead>
<tbody>
<tr>
<td>CMV infection</td>
<td>Infections other than CMV and bacterial:</td>
<td>GVHD</td>
</tr>
<tr>
<td>Clostridium difficile infections (CDI)</td>
<td>Viruses (adenovirus, astrovirus, human herpes virus 6/hhv-6, norovirus, dengue fever), fungi (candida), and parasites (blastocystis, hymenolepis nana, trichuristrichiura, isospora belli, and Entamoeba)</td>
<td>Non-infectious colitis (IBD)</td>
</tr>
<tr>
<td>Small bowel bacterial growth (SBBO): Escherichia coli, Campylobacter, Shigella, Salmonella</td>
<td></td>
<td>Drugs other than MMF</td>
</tr>
<tr>
<td>MMF drug therapy</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**Evidence Summary**

Due to frequent exposure to antibiotics and repeated hospitalizations SOT patients are a risk of developing intra-abdominal infections (IAI). IAI is commonly seen as a complication in post-liver, pancreas, multivisceral transplantation and superinfection with MDR pathogens occur frequently causing tertiary peritonitis. Common causes of diarrhea are as mentioned in Table 3. The treatment of choice for CDI had been oral vancomycin. Vancomycin has been shown to have much better efficacy compared to metronidazole in many studies. Zar et al in their randomized controlled trial in 150 patients found that both metronidazole and vancomycin were equally effective in treating mild CDI, but Vancomycin was much more effective in treating severe CDI. The clinical cure rate in mild CDI was 90% and 98% with metronidazole and vancomycin respectively (p = 0.36). In severe CDI the cure rates were 76% and 97% respectively (p = 0.02). Recurrence rates were also comparable in both the groups. Fekety et al in their randomized controlled trial compared two doses regimen of vancomycin in 46 patients viz. 125 mg four times a day vs. 500 mg four times a day. They found no difference in measurable responses to the two regimens. Since 125 mg four times a day is more cost-effective 125 mg dose is recommended. Even 125 mg-dose produces stool concentration of vancomycin of around 100 times more than the minimal inhibitory concentration (MIC) for C. difficile. The usual dose of oral vancomycin for children is 40mg/kg daily given in three or four divided doses. Fecal microbiota transplantation (FMT) in the management of refractory CDI has gained popularity recently, although in SOT recipients it has theoretical safety concerns. A series of cases (75 adults and 5 pediatric patients) treated with FMT for recurrent, refractory, and severe and/or overlap of recurrent/refractory and severe CDI had 78% cure rate after first FMT. There were no related infectious complications or adverse events in these high-risk patients.

**CNS Infections**

**Evidence Statements**

- We recommend initial workup for suspected CNS infections should include (I, A)
  a. MRI over CT scan
  b. CSF analysis including India ink preparation
  c. Rapid multiplex PCR on CSF
  d. Serum cryptococcal antigen
- We recommend empiric treatment to be started with Ceftriaxone + Vancomycin + Acyclovir (I, A)
- We recommend Liposomal Amphotericin B as the initial treatment for Cryptococcus. (I, A)

**Evidence Summary**

SOT patients with altered sensorium should be evaluated with detailed workup. Multifactorial etiologies coexist which are often obscured in these group of patients. Common pathogens causing CNS infections in SOT recipients are mentioned in Table 4. Although each imaging modality has unique insight to diagnose pathophysiology, but magnetic resonance imaging (MRI) is the preferred modality. It can diagnose infectious as well as non-infectious etiologies like drug toxicities, metabolic disorders as well as the progression of the disease and response to the therapy. Empiric broad-spectrum antimicrobial therapy including viral and fungal infections are preferred. It is preferred to use empirical bactericidal or fungicidal agents having CNS penetration until a diagnosis is achieved. There has been always a risk of donor-derived infections in SOT recipients thus donors should be screened with standard screening tests.
Nocardia: Post transplantation

- We recommend the following regimens for treatment of post-transplant nocardia infections
  1. **Pulmonary**: TMP-SMX (I, A)
  2. **Disseminated or CNS, Critically ill**: Imipenem plus TMP-SMX or Amikacin (I, A)
  3. **Alternative**: Meropenem, Linezolid (I, A)

Evidence Summary

SOT recipients are at risk of developing nocardia infection which is an opportunistic event. The risk of developing nocardiosis after SOT varies with the type of organ transplanted, e.g., the highest incidence in recipients of a lung transplant. A review of 5126 organ transplant recipients has demonstrated that highest nocardial infection rate among lung transplant recipients (3.5%). TMP-SMX is the treatment of choice for nocardial infections as it has demonstrated clinical efficacy and achieves high tissue concentrations in lung, brain, skin, and bone. Combination therapy is recommended in critically ill patients with pulmonary nocardia, cerebral nocardia, and disseminated nocardia. Linezolid has shown good activity against all species of nocardia.

REFERENCES


PART 3. THE HUMAN IMMUNODEFICIENCY VIRUS (HIV) POSITIVE PATIENT IN THE INTENSIVE CARE UNIT

These guidelines are applicable to a patient who is known to be infected with the human immunodeficiency virus (HIV) or presents for the first time to ICU with AIDS-defining conditions.

With the advent of highly active anti-retroviral therapy (HAART) era and the Test and Treat policy in HIV (where anyone testing positive receives HAART irrespective of CD4 cell counts), the incidence of ICU admissions for an HIV related illness is decreasing. Although HIV infected patients may still seek intensive care for reasons related directly to HIV infection, more and more seek care for other conditions that are unrelated to HIV such as trauma, infections, and other chronic diseases. Except for some special conditions such as opportunistic infections or HIV related treatment complications, HIV-infected patients are managed similarly to other patients without HIV infection. The treating physician should be aware of drug interactions, infectious and noninfectious conditions as a cause of the clinical presentation.1

The HIV patient in ICU with Acute respiratory failure

Evidence statements

- Appropriate samples should be collected for staining and cultures—including sputum/induced sputum and bronchoscopic lavage—if indicated. (UPP, A)
- Patients with severe pneumonia who require intensive care and without risk of Pseudomonas aeruginosa should be empirically treated with an IV β-lactam plus IV macrolide (II, A). Preferred β-lactams are ceftriaxone, cefotaxime, or amoxicillin-clavulanic acid. In patients who are allergic to penicillin, aztreonam plus azithromycin should be used (III, B).
- Those with CD4 counts < 200/mm³ and without signs of focal consolidation may be suspected to have PCP and should receive Trimethoprim-sulphamethoxazole (Co-trimoxazole) in therapeutic dose (TMP 15–20 mg/kg/day plus SMX 75 to 100 mg/kg/day given q6h or q8h). (I, A)
- Patients with documented or suspected Pneumocystis Jerovceii pneumonia (PCP) pneumonia and moderate-to-severe disease, defined by room air PO₂ < 70 mm Hg or Alveolar-arterial O₂ gradient ≥ 35 mmHg, should receive adjunctive corticosteroids as early as possible and certainly within 72 hours after starting specific PCP therapy (I, A)
- If patients with HIV/AIDS develop acute respiratory failure and they have any of the risk factors for Pseudomonas infection we recommend dual antipseudomonal coverage such as anti-pseudomonal β-lactam plus aminoglycoside (examples of anti-pseudomonal β-lactams include ceftazidime, cefoperazone, cefoperazone-sulbactam, piperacillin-tazobactam, imipenem-cilastatin, or meropenem (III, A). In patients who are allergic to penicillin, aztreonam can be used in place of the β-lactam. Combination therapy may be considered with the addition of aminoglycosides or antipseudomonal fluoroquinolones (e.g., levofloxacin, ciprofloxacin) (III, B).
- We recommend continuing Azithromycin along with anti-pseudomonal therapy for coverage of atypical pathogens (II, B).
- We recommend against using fluoroquinolones empirically to avoid development of drug-resistant TB. Patients should also undergo sputum testing for acid-fast bacilli simultaneously if fluoroquinolones are being used. Fluoroquinolones may be continued only if tuberculosis is not a diagnostic consideration at admission (I, A).
- In patients who have risk factors for methicillin-resistant Staphylococcus aureus (MRSA) infection—empiric treatment should include vancomycin or linezolid (III, B).
- We suggest the addition of clindamycin (to vancomycin, but not to linezolid) in cases of severe necrotizing pneumonia to minimize bacterial toxin production (III, B).
- When the etiology of pneumonia has been identified on the basis of reliable microbiological methods, antimicrobial therapy should be de-escalated (II, A).
- A switch to oral therapy should be considered in patients with community acquired pneumonia (CAP) on IV antibiotic therapy who have improved clinically, can swallow and tolerate oral medications, and have intact gastrointestinal function (II, A).

Evidence Summary

Respiratory failure is the most important cause of ICU admission among HIV patients. However, the specific microbiology data on etiology among HIV positive patients in the ICU is lacking from the Indian subcontinent. After going through the available literature, the most common pathogens seem to be viral infections, Pneumocystis jirovecii, Streptococcal pneumonias, H. Influenzae, M. tuberculosis, Staphylococcus aureus, Klebsiella pneumonias, Pseudomonas aeruginosa, and Escherichia coli² (Table 1). These patients are highly susceptible to infections with Mycobacterium tuberculosis (MTB). Hence MTB should be actively searched and ruled out in this population.
and this may often require invasive interventions like bronchoscopy to get a bronchoalveolar sample. Appropriate sample collection may be taken for staining and cultures (sputum/bronchoscopic lavage or non-directed BAL in intubated patients Table 2). Sample collection should be done as early as possible, preferably within 1 hour and a broad spectrum antibiotic to cover gram-negative organisms may be added empirically. In case there is a delay of collecting the sample, the antibiotics should be administered as fast as possible. Sputum samples should be sent for gram staining, culture, and also special stains for pneumocystis. Cartridge based nucleic acid amplification test (CBNAAT) of samples is beneficial in the early and rapid diagnosis of MTB and multi-drug resistant tuberculosis (MDRTB). In concordance with the surviving sepsis guidelines, we also recommend measuring lactate at baseline.

**Table 3: Risk factors for Pseudomonas and Staphylococcal infections**

<table>
<thead>
<tr>
<th>Risk factors</th>
<th>Risk factors for methicillin-resistant S. aureus</th>
</tr>
</thead>
<tbody>
<tr>
<td>Advanced immunosuppression/full blown AIDS (CD4 count ≤50 cells/mm³)</td>
<td>Recent influenza infection; IV drug abusers</td>
</tr>
<tr>
<td>Underlying structural lung disease such as bronchiectasis</td>
<td>Severe, bilateral, necrotizing pneumonia</td>
</tr>
<tr>
<td>Profound neutropenia</td>
<td>Patients on chronic hemodialysis</td>
</tr>
<tr>
<td>Treatment with long-term corticosteroids</td>
<td>Patients on chronic hemodialysis</td>
</tr>
<tr>
<td>Severely malnourished patients</td>
<td></td>
</tr>
</tbody>
</table>
unknown efficacy of monotherapy, the expert committee felt that these patients should receive broad-spectrum combination therapy rather than monotherapy.

After starting on initial broad-spectrum antibiotics, once the culture reports are available, the antibiotic therapy should be de-escalated as quickly as possible. Antibiotic de-escalation reduces the possibility of adverse drug effects, treatment costs and reduces the incidence of drug resistance. Parenteral antibiotics can be changed to oral route once clinical stability is attained. Suggested criteria for clinical stability include oral temperature <37.8°C, heart rate <100 beats/minute, respiratory rate <24 breaths/minute, systolic blood pressure ≥90 mm Hg, and room air oxygen saturation >90% or partial pressure of oxygen in arterial blood (PaO₂) > 60 mm Hg. De-escalation is safe in patients hospitalized with CAP including severe CAP. Antibiotics need to be given for a minimum of 5 days of treatment and may be stopped in case patients remain afebrile for 48 to 72 hours and are clinically stable. A longer duration of treatment might be required in case of metastatic foci of infection, or infection with drug-resistant bacteria such as MRSA or drug-resistant gram-negative bacteria.

**HIV Positive Patients Presenting with Suspected Bloodstream Infections or Sepsis of Unknown Origin**

**Evidence Statements**

- In the presence of sepsis or septic shock, we recommend following the surviving sepsis guidelines similar to the management of other patients with sepsis (UPP, A).
- In the absence of septic shock or absence of risk factors for Pseudomonas a monotherapy with a third-generation cephalosporin or a cephalosporin, the β-lactamase inhibitor is sufficient (II, A)
- In more severe disease states, such as in the presence of organ dysfunction or septic shock—a combination of broad-spectrum antibiotics may be used for initial empiric therapy (III, A).
- Combination therapy is discouraged in the absence of ongoing shock (I, B)
- Empiric gram-positive coverage is suggested for those who have risk factors for MRSA (UPP, A)
- Anti-fungal agents may be considered only if there is no clinical improvement or there is clinical deterioration even after 72 hours of appropriate empirical antibiotics therapy and CD4 counts <200/mm³ (II, A).
- We recommend against the use of empirical antifungal therapy (II, A)
- Those with CD4 <200/mm³ are at high risk for disseminated tuberculosis and hence, need to be worked up for tuberculosis — including blood cultures for tuberculosis (I, B).
- We recommend against empirical anti-tubercular therapy (ATT). In cases of proven mycobacteremia, ATT may be started as per national guidelines or in consult with the ID specialist (I, A).

**Table 4:** Results of comparison of various antibiotic regimen in Community Acquired Pneumonia.

<table>
<thead>
<tr>
<th>Author/ Country</th>
<th>Design</th>
<th>Study population</th>
<th>Conclusion</th>
</tr>
</thead>
<tbody>
<tr>
<td>Postma DF, Netherlands ⁸</td>
<td>randomized, crossover trial</td>
<td>Hospitalized patients (non-ICU)</td>
<td>Among patients with clinically suspected CAP, empirical treatment with beta-lactam monotherapy was non-inferior to a beta-lactam–macrolide combination or fluoroquinolone monotherapy with regard to 90-day mortality.</td>
</tr>
<tr>
<td>Lee JS ⁹</td>
<td>Review</td>
<td>adults hospitalized with community-acquired pneumonia</td>
<td>A cluster randomized trial demonstrated an absolute adjusted difference of 2.5% in 90-day mortality favoring β-lactam monotherapy over β-lactam plus macrolide combination therapy. Another randomized trial found an absolute difference of 7.6% in the attainment of clinical stability on day 7 favoring β-lactam plus macrolide combination therapy over monotherapy with β-lactam. Six observational studies found that β-lactam plus macrolide combination therapy was associated with relative reductions of 26% to 68% in short-term mortality. 3 observational studies reported that fluoroquinolone monotherapy was associated with relative reductions of 30% to 43% in mortality compared with β-lactam monotherapy.</td>
</tr>
<tr>
<td>Nie W ¹⁰</td>
<td>Meta-analysis</td>
<td>Included four prospective cohort studies and 12 retrospective cohort studies</td>
<td>In comparison with beta-lactam monotherapy, beta-lactam macrolide dual therapy reduces the risk of mortality in patients with CAP.</td>
</tr>
<tr>
<td>Garin N, Switzerland ¹¹</td>
<td>multicenter, non blinded, noninferior, RCT</td>
<td>580 immunocompetent adult patients with moderately severe CAP</td>
<td>In patients hospitalized for moderately severe CAP, β-lactam monotherapy had delayed clinical stability with monotherapy and hence cannot be considered non inferior.</td>
</tr>
</tbody>
</table>
• Lateral Flow urine LipoArabinomannan Assay (LF-LAM) may be used to assist in the diagnosis of TB only in HIV positive adult in-patients with signs and symptoms of TB (pulmonary and/or extrapulmonary) who have a CD4 cell count < 100 cells/mm³, or HIV positive patients who are seriously ill regardless of CD4 count or with unknown CD4 count (II, B).
• LF-LAM should not be used as a screening test for TB (III, A).

Evidence Summary

Bloodstream infections are common in AIDS patients. As with failure cases, most of the data is from western countries (Table 5). Except for blood stream infections (BSI) caused by tuberculosis, the data from India is scarce. The common organisms seem to be non-typhoid Salmonellae, Streptococcus pneumoniae, Escherichia coli, Staphylococcus aureus, and coagulase-negative Staphylococci. M. Tuberculosis is not rare if CD4 count < 100/mm³. The antibiotics policy for these patients remains the same as mentioned in international guidelines for sepsis.17,18

HIV positive patient presenting with signs of CNS infection in ICU

Evidence Statements

• We recommend brain imaging (preferably MRI) and/or fundoscopy for ruling out raised intra cranial pressure (ICP). If possible the opening pressure should be measured, and physical characteristics such as cerebrospinal fluid (CSF) appearance should be noted (UPP, B)
• A lumbar puncture should be performed for microbiological and biochemical assay including gram stain, India ink stain, aerobic culture, protein, sugar, Adenosine Deaminase (ADA) levels, lactate levels, and cells.
• We suggest a detailed workup for Tuberculosis including an assay for CBNAAT, Mycobacteria growth indicator tube (MGIT) culture, CSF analysis, ADA. (UPP, B)
• In immunocompromised host with relevant clinico-radiological findings and CNS symptoms, additional samples may be sent for T. gondii (IgG antigen and/or polymerase chain reaction (PCR), cryptococcal antigen, Mycobacterium tuberculosis (PCR), and common viruses such as Epstein-Barr Virus DNA (EBV) and JC Virus (JCV) along with work up for bacterial culture (UPP, A).

Table 5: Common organisms isolated from the bloodstream in patients with HIV

<table>
<thead>
<tr>
<th>Reference (First Author, Year)</th>
<th>Study Location and Time Frame</th>
<th>Study Design</th>
<th>Primary Inclusion Criteria</th>
<th>Main Isolates in HIV Patients (% of Bacterial Isolates in HIV)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Grant, 199719</td>
<td>Ivory Coast, 1995</td>
<td>Prospective Observational Study</td>
<td>Admission to the infectious disease unit</td>
<td>Nontyphoid salmonella 23 (59), E. coli 6 (15), S. pneumoniae 4 (10)</td>
</tr>
<tr>
<td>Hung, 199820</td>
<td>Taiwan, 1994–1996</td>
<td>Prospective Observational Study</td>
<td>Fever (≥38°C) in HIV-infected patients admitted to AIDS unit</td>
<td>Non typhoid salmonella 24 (80), S. aureus 3 (8), Klebsiella pneumoniae 2 (5), E. coli 2 (5), P. aeruginosa 2 (5)</td>
</tr>
<tr>
<td>Manfredi, 199921</td>
<td>Italy, 1994–1995, 1997–1998</td>
<td>Retrospective study</td>
<td>HIV patients admitted to the Hospital</td>
<td>Gram-positive catalase-positive cocci S. pneumoniae 68 (43), NTS 42 (26), CoNS 10 (6), E. coli 9 (6)</td>
</tr>
<tr>
<td>Mayanja, 201022</td>
<td>Rural Uganda, 1996–2007</td>
<td>Cohort study</td>
<td>Fever (≥38°C) with no detectable malaria Parasites</td>
<td>Nontyphoid salmonella 12 (75)</td>
</tr>
<tr>
<td>Meremo, 201223</td>
<td>Urban Tanzania, 2011</td>
<td>Prospective Observational Study</td>
<td>Hospital admission with fever (axillary temperature &gt;37.5° C)</td>
<td>NTS 82 (61), S. aureus 13 (20), Enterobacter spp 11 (8)</td>
</tr>
<tr>
<td>Mootsikapun, 200724</td>
<td>Urban Thailand, 1996–2001</td>
<td>Retrospective study</td>
<td>Positive blood culture (including fungi and mycobacteria)</td>
<td>NTS 82 (61), S. aureus 13 (20), Enterobacter spp 11 (8)</td>
</tr>
<tr>
<td>Muyanja, 201125</td>
<td>Uganda, 1995–2008</td>
<td>Cohort study</td>
<td>HIV infected</td>
<td>S. pneumoniae 103 (42), NTS 66 (27), E. coli 18 (7)</td>
</tr>
<tr>
<td>VarmaJK, 201026</td>
<td>Thailand, 2006–2008</td>
<td>Prospective–2006–2008</td>
<td>Outpatients diagnosed with HIV regardless of the presence or absence of symptoms or prior suspicion of clinical illness</td>
<td>M.TB 31 (54%), fungi [13 (22%)], and bacteria [9 (16%)].</td>
</tr>
<tr>
<td>Gopinath27</td>
<td>India, 2005–2006</td>
<td>Prospective study</td>
<td>Hospitalized patients with HIV-and suspected tuberculosis</td>
<td>M.TB 30%</td>
</tr>
<tr>
<td>Ramachandran28</td>
<td>India</td>
<td>Prospective control study</td>
<td>Hospital admission with a diagnosis of HIV</td>
<td>4% MTB</td>
</tr>
</tbody>
</table>
For Cryptococcal Meningitis, a paired sample of both serum and CSF should be sent for S. Cryptococcal antigen assay and CSF Cryptococcal antigen (UPP, A).

In patients presenting with features of raised ICP and intracranial mass lesions, toxoplasma also needs to be considered. We recommend CSF analysis for IgG antibodies against Toxoplasma antibodies as it has a good negative predictive value in ruling out infections (UPP, A).

For a patient coming to ICU with altered CNS function and suspicion of meningitis, we recommend a third-generation cephalosporin known to penetrate the blood-brain barrier - at higher doses, e.g., Ceftriaxone 2 gm BD intravenously (I, A).

We recommend the addition of vancomycin empirically to the initial treatment regime (I, B).

We recommend de-escalating antibiotics after culture reports are available (I, A).

In patients above 50 years of age, we suggest the use of additional ampicillin at high doses of 2 gm every 6th hourly (I, B).

In very young infants of age < 1 month, we suggest Ampicillin plus cefotaxime or ampicillin plus an aminoglycoside as the initial management (I, B).

For Cryptococcal meningitis, we recommend induction therapy followed by consolidation therapy for 2 months. In patients with severe disease, and low CD4 counts we recommend maintenance therapy for at least a year (II, A).

The preferred induction therapy recommended is Liposomal amphotericin B (3 to 5 mg/kg IV daily) plus flucytosine (100 mg/kg per day orally in four divided doses) for a minimum of two weeks. If flucytosine is not available, fluconazole (800 mg daily orally) with Amphotericin B should be given for a minimum of two weeks (I, B).

In view of the risk of serious adverse effects, we suggest the use of liposomal amphotericin B instead of amphotericin deoxycholate, if cost is not an issue (II, B).

We recommend Amphotericin B deoxycholate (0.7 - 1 mg/kg/day IV daily) be used if liposomal amphotericin is not available and the risk of renal dysfunction is low. (I, A)

Consolidation therapy with fluconazole at a dose of 400 mg orally daily for a minimum of eight weeks is recommended (I, A).

Maintenance therapy: At the completion of eight weeks, fluconazole 200 mg daily should be continued for long-term suppression for a minimum of one year, if CD4 count >100 cells/mm³ (II, B).

Secondary prophylaxis should be reinitiated if the CD4 count decreases again to <100 cells/mm³ (III, A).

Corticosteroids and mannitol have been shown to be ineffective in managing ICP, and we recommend against the routine use of these agents in cryptococcal meningitis (III, A). In the case of IRIS, corticosteroids should be administered to manage severe central nervous system immune reconstitution inflammatory syndromes. (I, A)

Immediate ART in the setting of Cryptococcal meningitis may increase the risk of serious IRIS, a short delay (4–8 weeks) before initiating ART may be necessary.²⁹,³⁰

Empiric therapy for suspected CNS toxoplasmosis

Evidence Statements

We recommend a combination of pyrimethamine plus sulfadiazine plus leucovorin in patients with suspected toxoplasmosis (Sulfadiazine 1000 mg four times daily among patients <60 kg or 1500 mg four times daily among patients ≥60 kg, Pyrimethamine 200 mg loading dose followed by 50 mg daily among patients <60 kg or 75 mg daily among patients ≥60 kg, Leucovorin 10 to 25 mg daily) (I, A).

Pyrimethamine plus clindamycin plus leucovorin is the preferred alternative regimen for patients with TE who cannot tolerate sulfadiazine or do not respond to first-line therapy (I, A).

If pyrimethamine is unavailable or there is a delay in obtaining it, TMP-SMX should be utilized in place of pyrimethamine-sulfadiazine or pyrimethamine-clindamycin (I, B).

Acute therapy for Toxoplasmosis should be continued for at least 6 weeks if there is a clinical and radiologic improvement (II, B).

Adjunctive corticosteroids such as dexamethasone may be administered to patients to treat a mass effect associated with focal lesions or associated edema (III, B)

Management of Tuberculous meningitis or CNS Tuberculoma

The management remains the same as in CNS tuberculosis in an immunocompetent individual with 2 months of an intensive regimen with four drugs and 7 to 9 months of continuation phase (total 9–11 months) (I, A).

If not already on ART, ART should be initiated after 8 weeks of intensive phase, regardless of CD4 count (I, A).
Drug interactions need to be considered when starting treatment of an HIV patient already on HAART with drugs such as rifampicin and clarithromycin. Drug resistance is an important factor that has to be considered in these patients hence whenever possible, the expert committee feels that treatment of suspected TB should be initiated only after a microbiological diagnosis and in liaison with an infectious disease expert.

**Evidence Summary**

Management of treatment-naïve HIV-infected patients with TB is especially challenging in areas with high rates of coinfection. Initiation of antiretroviral therapy may be complicated by the immune reconstitution inflammatory syndrome (IRIS), which can manifest as reactivation of latent TB, the progression of active TB disease, or clinical deterioration in patients previously improving on antituberculous therapy.

The most important factor in determining the differential diagnosis is the degree of immunosuppression in the patient.

In patients with CD4 cell counts > 500/µm³, benign and malignant brain tumors and metastases predominate, as in immunocompetent hosts.

In moderately immunosuppressed patients with CD4 cell counts from 200 to 500/µm³, HIV-associated cognitive and motor disorders are common but usually, do not present with focal lesions.

In severely immunosuppressed patients with CD4 cell counts < 200/µm³, CNS mass lesions are most common. The most likely diagnostic considerations include opportunistic infections (Toxoplasma encephalitis, primary CNS lymphoma, progressive multifocal leukoencephalopathy, HIV encephalopathy, and CMV encephalitis) and AIDS-associated tumors, such as primary central nervous system lymphoma.

In addition, multiple etiologies can coexist in an immunosuppressed individual.31-33

Patients who are not under medical supervision, or those who are not aware of their HIV status, may present with their first opportunistic infection in the central nervous system. The clinical manifestations and the diagnostic possibilities are similar to those seen in before the introduction of potent antiretroviral therapy (ART) era. However, the effect of prophylaxis for Pneumocystis and the use of ART may alter the clinical spectrum of disease and the diagnostic considerations in patients who are taking medications.34-35 In a patient presenting with CNS symptoms/ altered mentation, noninfectious and infectious causes need to be ruled out. A practical approach is shown in the flowchart.


Evidence Statements

- If an asplenic/hyposplenic patient is suspected to have sepsis, we recommend administration of IV ceftriaxone before transferring the patient to a higher center (II, A).
- We recommend that all patients with overwhelming post-splenectomy infection (OPSI) be treated in the ICU (UPP, A).
- We recommend empiric antibiotic therapy for asplenic patients with a combination of ceftriaxone and vancomycin (I, A).
- In case of allergy to β-lactams, we recommend vancomycin with aztreonam or fluoroquinolones in adults. Do not delay the administration of antibiotics, be prepared to treat the reaction (UPP, A).
- We recommend adding clarithromycin or erythromycin in case of respiratory symptoms (III, A).
- We recommend empiric therapy with IV Cefotaxime + vancomycin + ampicillin if the patient age < 2 months: (III, A).
- All febrile asplenic patients should be screened for malaria with peripheral smears. Start artesunate-based antimalarial therapy, if the history is suggestive of Malaria (UPP, A).
- If gram staining of peripheral blood smear shows gram-negative bacilli, we recommend the addition of antipseudomonal coverage to the therapy (III, A).
- We recommend that urine be checked for a urinary antigen for streptococcus pneumonia (II, A).
- We suggest RT-PCR test for simultaneous identification of 3 main encapsulated bacteria (Strep pneumonia, H. influenzae type B and N. meningitidis) (III, B).
- We recommend that all asplenic patients should receive immunization against encapsulated bacteria (S. pneumoniae, H. influenzae, and N. meningitidis) (I, A).
- Immunization against seasonal flu is recommended for patients over 6 months of age (I, A).
- We recommend that vaccination programs should be started no sooner than 14 days after splenectomy (I, A).
- If the patient is discharged before 15 days after splenectomy or angioembolization, where the risk to miss vaccination is deemed high, we suggest that patient be vaccinated before discharge (I, B).
- Antibiotic prophylaxis is indicated in patients for 1-2 years after splenectomy and lifelong for the patient had an episode of overwhelming infection or immunocompromised (II, B).
- We recommend self-administration of one dose of, in stock “pill in the pocket”, prescribed antibiotics in the event of any sudden onset of unexplained fever, malaise, chills or other constitutional symptoms, when medical consultation not readily accessible within 2 hours (II, A).
- We suggest that any patient with sepsis having a risk factor for hyposplenia, the peripheral smear should be checked for Howell-Jolly bodies (II, B).
- We recommend formulation of Spleen registry (UPP, A).

Evidence Summary

Splenectomy is often indicated in patients with an underlying malignant or nonmalignant hematologic diseases or cases of splenic rupture following infection or trauma. Other causes of hyposplenia include auto infarction in subjects with sickle cell anemia and chronic graft-versus-host disease after stem-cell transplantation, severe celiac disease, and untreated human immunodeficiency virus infection.5

Overwhelming post-splenectomy infection (OPSI) is defined as an infection, occurring more commonly after splenectomy (or in hyposplenic host) which evolves over a short time and produces severe symptoms, often with hypotension and a high mortality rate.6

Patients with hyposplenism due to splenectomy or hyposplenia are at an increased risk for invasive infections with encapsulated bacteria as Streptococcus pneumoniae, Haemophilus influenza type b, Neisseria meningitidis.2,7 These infections progress rapidly from a mild flu-like illness to fulminant sepsis and are associated with a high mortality rate of up to 50% despite maximal treatment.

PART 4. INDIAN ANTIMICROBIAL PRESCRIPTION GUIDELINES FOR PATIENTS WITH CONGENITAL AND ACQUIRED HYPOSPLENISM/ASPLENIA

Patients with congenital and acquired hyposplenism/asplenia are prone to specific infections and are at increased risk of severe sepsis. Although the incidence of septicemia remains low, the risk for overwhelming post-splenectomy infection (OPSI) remains higher than in the general population.1 Majority of the overwhelming post-splenectomy infection (OPSI) is caused by encapsulated bacteria such as Streptococcus pneumoniae, Haemophilus influenza, and Neisseria meningitides although other infections can also occur.2-3 The disease starts as a minor flu-like illness and rapidly evolves into a fulminant course of hypoglycemia, metabolic acidosis, dyselectrolytemia, disseminated intravascular coagulation (DIC), shock, coma and death within 24 to 48 hours.4 OPSI usually occurs within the first two years after splenectomy but may also occur later and has a mortality rate of 50 to 70% despite aggressive therapy.2 In view of the severe progression and high mortality of OPSI, stress has been given for early aggressive treatment as well as immunization of patients with splenectomy and thereby preventing OPSI.
The lifetime risk of OPSI is assumed to be 5%, and the highest frequency of these OPSIs is during the first 2 years after splenectomy.\(^6\) Patients with sickle cell anemia, thalassemia major or malignancies such as Hodgkin’s lymphomas and non-Hodgkin’s lymphomas have a higher risk for OPSI. Asplenic patients have a higher incidence of parasitemia, a delayed clearance of parasites after treatment or a severe or even fatal infection due to malaria. These patients are also at high risk for Babesiosis, and this might be confused with *Plasmodium falciparum*.\(^2\) These patients are at an increased risk of OPSI with Capnocytophaga canimorsus, if bitten by dogs and other animals and should receive adequate antibiotic coverage following such bites.\(^10\) Otherwise rare, Ehrlichiosis is also more severe in patients with asplenia/hyposplenia.\(^2\)

OPSI should be considered as a medical emergency and mandates early recognition and aggressive management. These patients should be managed aggressively including immediate cultures and administration of a combination of antibiotics to cover all possible etiological agents. In areas where penicillin-resistant pneumococci are prevalent, other agents such as vancomycin, teicoplanin or rifampicin should be added to ceftriaxone as the initial empiric therapy. Gram stain of the peripheral blood or buffy coat will give an idea regarding the presence or absence of intraleukocytic bacteria. Antipseudomonal coverage should be added in case of high risk for Pseudomonas infection or peripheral blood growing GNB. The presence of intracellular bacteria within leukocytes should alert the clinician towards ehrlichiosis while the presence of parasites in RBC should alert for malaria or babesiosis. Once the blood cultures are positive antibiotics can be modified accordingly.

**REFERENCES**

Primary immunodeficiencies are a group of disorders that affect the development, function or both of the immune system. There are more than 300 disorders defined till date. The prevalence is approximately 1 in 10,000 live births. Any patient admitted to ICU could be a potential PID patient.

**Diagnosis of PID**

**Evidence Statements**

- PID should be suspected when the following history/symptoms or signs are present (UPP, A)
  1. Family history of sibling death
  2. Four or more ear infections within 1 year
  3. Two or more serious sinus infections or pneumonias within 1 year
  4. Two or more months on antibiotics with little effect
  5. Two or more deep-seated infections including septicemia
  6. Persistent thrush in mouth or fungal infection on the skin
  7. Infections in multiple anatomic locations
  8. Increasing frequency and severity of infections with age
  9. Recurrent serious infections with common pathogens
  10. Serious infections with unusual pathogens

- We recommend that when PID is suspected, HIV infection should also be considered, and testing should be performed for HIV (UPP, A)

- We recommend that patient should be investigated for PID when:
  1. In neonates, absolute lymphocyte count (ALC) of < 2000/mm³ in cord blood or an infant an ALC of < 4000/mm³
  2. Severe hypogammaglobulinemia with IgG < 150mg/dL
  3. Absolute lymphocyte count < 4000/mm³ (In non-chemotherapy setting)
  4. Unusual organism picked up on microbiology
  5. Unexplained neutropenia

- We recommend that Initial laboratory screening should include a complete blood count with differential counts (including absolute lymphocyte count, absolute neutrophil count, absolute monocyte count) and measurement of serum immunoglobulin and complement levels (UPP, A). We recommend severe combined immune deficiency (SCID) be considered as a pediatric emergency and attention be paid to absolute lymphocyte count, at all time in ICU. If the absolute lymphocyte count is less than normal for the age, we recommend to make immunology reference, use irradiated blood products, and avoid live vaccines till diagnosis is confirmed or ruled out (UPP, A).

- We recommend that patient be investigated for combined variable immuno-deficiency (CVID) when the patient has any of the following: (UPP, A)
  1. Recurrent bacterial infections.
  2. Serum IgG, IgM, IgA levels (at least two of the three) with a marked decrease (at least 2 SD below the mean for age)
  3. The onset of immunodeficiency at more than 2 years of age.
  4. The absence of isohemagglutinins and or poor response to vaccines.

- We recommend that immunology consult be obtained for these patients and the patient be investigated to diagnose specific form of immunodeficiency (UPP, A)
  1. Lymphocyte subpopulations by flow cytometry (CD3, CD4, CD8, CD19, CD20, CD16 and CD56).
  2. Naive T cells, Memory B cells, Memory T cells
  3. T-cell response to mitogens.
  4. Nitroblue tetrazolium-NBT test
  5. Complement levels
  6. Bone marrow and genetic tests

**Evidence Summary**

Diagnosis is often delayed since signs and symptoms such as bronchitis, pneumonia, sinusitis, and diarrhea are considered infection related without suspecting immunological process.

The absence of adenoid tissue in the nasopharynx or absence of the thymus should prompt suspicion of primary immunodeficiency (antibody or cellular/com-bined).

The presence of lymphocytopenia on complete blood count suggests a T-cell disorder, whereas a finding of neutropenia suggests a phagocytic disorder. Abnormal serum immunoglobulin levels suggest a B-cell disorder. Abnormalities on assay of the classic or alternative complement pathways suggest a complement disorder.

Abnormal values of lymphocyte count should also raise suspicion of PID (Table 1).

**Evidence Statement**

- We recommend appropriate cultures and PCRs; for organisms likely to cause infections pertinent to the conditions they are suffering from (UPP, A).
- An attempt should be made to identify the microorganisms directly or on PCRs as serological tests in infectious diseases could give false-negative results if there is an antibody defect (UPP, A).
- We recommend the use of Multiplex PCR to help diagnose infections (UPP, A)
Evidence Summary

Patients with PID commonly present with recurrent infections and invasive infections, atypical pathogens, partial response to antibiotics, failure to thrive, chronic diarrhea, fungal infections, unexplained skin rash, and family history. Infections such as Pneumonia and bronchiolitis, acute gastroenteritis, otitis media, and bacteremia in patients with an antibody, combined, and cellular deficiencies. Whereas viral infections meningitis, osteomyelitis, gastroenteritis is commonly seen in CVID. Children tend to have bacterial or fungal infections with unusual organisms or unusually severe and recurrent infections with common organisms. A family history of primary immunodeficiency disease is the strongest predictor of a person having this type of disease.5

The typical presentations of various PIDs by age of presentation and spectrum of infections.

- Combined T-cell and B-cell immunodeficiency (Presents early in life)
  1. Bacteria: Campylobacter Listeria, Pyogenic bacteria, Mycobacteria
  2. Viruses: RSV, EBV, Parainfluenza Virus
  3. Fungi: Candida, Aspergillus
  4. Protozoa: Pneumocystis jiroveci, Toxoplasma Gondi, Cryptosporidium parvum

- B cell immunodeficiency (Presents when weaning is started and breastfeeding stops)
  1. Bacteria: S. pneumonia, H. influenza, M. catarrhalis, P. aeruginosa, S. aureus, N. meningitidis, M. pneumonia
  2. Viruses: Enteroviruses
  3. Protozoa: Giardia lamblia

- Congenital defects of phagocyte number and function (Can present at any age based on the severity of the defect)
  1. Bacteria: S. aureus, P. aeruginosa, Nocardia, S. Typhi
  2. Fungi: Candida, Aspergillus
  3. Mycobacteria: Nontuberculous including BCG

- Complement deficiencies (Can present as early as within 6 months of life)
  1. Bacteria: Streptococci, H. influenza, Neisseria,
  2. Viruses: CMV, HSV

The European Society for Immunodeficiency (ESID) clinical guidelines proposed the grouping of immunodeficiency, syndromes and likely infections as follows (Table 2).

<table>
<thead>
<tr>
<th>Immunodeficiency</th>
<th>Infections</th>
<th>Example</th>
</tr>
</thead>
<tbody>
<tr>
<td>Antibody deficiency</td>
<td>Bronchiectasis, rhinosinusitis</td>
<td>HIV, Wiskott-Aldrich Syndrome</td>
</tr>
<tr>
<td>Neutrophil deficiency</td>
<td>Chronic pyogenic infections, Invasive Aspergillus, Burkholderia</td>
<td>Chronic granulomatous disease (CGD)</td>
</tr>
<tr>
<td>Defects of innate immunity (TLR3)</td>
<td>Invasive pneumococcal disease</td>
<td>SCID/HIV, Wiskott-Aldrich Syndrome</td>
</tr>
<tr>
<td>T lymphocyte/macrophage deficiency</td>
<td>Meningococci, encapsulated bacteria or Candida</td>
<td>Mycobacteria</td>
</tr>
<tr>
<td>Common variable immunodeficiency (CVID)</td>
<td>Autoimmune or chronic inflammatory disease</td>
<td>Haemophagocytic lymphohistiocytosis (HLH)</td>
</tr>
</tbody>
</table>

In ICU setting in patients with PID; following organisms are likely to cause infections.

- B cell deficiency
  1. Pneumococcus
  2. H. influenza
  3. Staph Aureus
  4. Giardia Lamblia
  5. Viruses Enterovirus/echovirus

- T cell deficiency
  1. Mycobacteria
  2. Viruses: CMV/EBV/HSV/RSV/VZ/ Parainfluenza
  3. Fungi: P.carini, Histoplasma, cryptosporidium, Toxoplasma

- Phagocytic disorder
  1. Gram Negative: E.coli/ Klebsiella/ B.cepecia/ Pseudomonas/ Serratia
  2. Gram positive: Staph /Nocardia/ Listeria/
  3. Fungi: Aspergillus and candida

- Defects in the complement system: Streptococcus pneumonia and Neisseria
• Mendelian susceptibility to Mycobacterium (MSMD): Mycobacteria, salmonella typhi, and nontyphi, listeria, viral and other intracellular pathogens (e.g., Histoplasma, leishmania).5-11
• We recommend intravenous immunoglobulin for treatment of all antibody deficiency diseases (UPP, A). The recommended dose is 400 mg/kg/doses every 4 weekly. We recommend 2 gm/kg single dose (severe infections) or 1 gm/kg weekly till infection subsides (UPP, A).
• We recommend to maintain serum IgG trough levels above 500 mg/dL and above 700 mg/dL in bronchiectasis (Level III, A).
• We recommend a computed axial tomography, lung function tests with spirometry and DLCO every 6 months after discharge (UPP, A).
• We recommend hematopoietic stem cell transplantation in cellular and macrophage immunodeficiency (UPP, A).
• We recommend monoclonal antibodies such as rituximab only in autoimmune complications related to CVID (UPP, A).
• We recommend Rituximab be given in PID complicated with EBV viremia (UPP, A).
• We recommend for all critically ill patients with suspicion of PID the empirical antimicrobial treatment with IV Carbapenems with IV Vancomycin/Teicoplanin for broad-spectrum coverage (UPP, A). Voriconazole is the preferred antifungal in case of proven, possible or probable invasive fungal infection with Aspergillus (I, A).
• In critically ill patients diagnosed with Combined B and T cell deficiency, the antimicrobial drug of choice is IV Carbapenems with IV Vancomycin/Teicoplanin and Trimethoprim-Sulfamethoxazole (UPP, A).
• In critically ill patients diagnosed with Combined B and T cell deficiency with suspicion of viral infections, we recommend: (UPP, A).
  1. IV Acyclovir if herpes group of infection is suspected
  2. Oral Oseltamivir if Influenza virus is suspected
  3. IV Ganciclovir if CMV is suspected radiologically or by laboratory tests
• In critically ill patients diagnosed with B cell deficiency, on the organisms expected (Capsulated), we recommend IV ceftriaxone with IV Vancomycin/Teicoplanin (UPP, A).
• We recommend IV immunoglobulin (IVIG) at a dose of 1 gm/kg weekly in cases of severe infections especially ECHO/Enterovirus/Polio virus-induced encephalitis (UPP, A).
• In critically ill patients diagnosed with Phagocyte disorder, we recommend antimicrobial drug of choice to be IV Carbapenems with IV Vancomycin/Teicoplanin and Voriconazole (UPP, A).
• We recommend the use of Granulocyte-colony stimulating factor (GCSF) in patients of congenital Neutropenia. (UPP, A)
• In critically ill patients diagnosed with complement deficiency, the antimicrobial drug of choice is IV Cephalosporin (UPP, A).

Evidence Summary
The data regarding the use of Antibiotics in Immunodeficiency states are scarce. The experts recommend using antibiotic as per organism isolated or expected. Generally, the management depends upon the type of PID.

Therapy includes
• IV Immunoglobulin (IVIG) infusion mainly for B cell deficiency.12-14
• Antibiotics as per suspected source of infection and suspected organism
• Rituximab in PID with Epstein Barr virus reactivation
• Stem cell transplant is the most curative option for the majority of the PID. Paradoxically Rituximab treatment has known to aggravate primary immunodeficiency or hypogammaglobulinemia in a certain group of patients, and appropriate care has to be taken in these patients.
• In PID such as X-linked Lymphoproliferative disorder, Rituximab can be given once in 4 weeks to decrease the EBV Viral load.1,3,4

Vaccinations and Antimicrobial Prophylaxis at Discharge

Evidence Statements
• We wish to emphasize that all forms of live vaccines, viral and bacterial, are contraindicated in patients with SCID (UPP, A).
• We recommend vaccination for diagnosed patients with complement deficiency at the time of discharge (UPP, A). We recommend to avoid BCG vaccination in chronic granulomatous disease/MSMD patient (UPP, A).
• We recommend antifungal, and anti PCP prophylaxis for all patients diagnosed with PID shifted from ICU. (UPP A). We recommend antifungal prophylaxis for all patients with Combined B and T or T cell deficiency with the drug of choice being Trimethoprim-Sulfamethoxazole (I, A) and PCP prophylaxis should be given to all patients with Combined B and T or T cell deficiency with the drug of choice being Trimethoprim-Sulfamethoxazole (I, A). We recommend antifungal prophylaxis in all patients with T cell defects (III, A).
Evidence Summary

Vaccine recommendations should be earmarked only for patients with certain PID.

Live vaccines are avoided in patients with severe B- and T-cell dysfunction due to the risk of dissemination and the futility of immune response. All vaccines are safe and effective in the patients with complement deficiency (susceptibility to encapsulated organisms).15-17

REFERENCES

1 Professor and Head, Division of Critical Care Medicine, Department of Anaesthesiology, Critical Care and Pain, Tata Memorial Hospital, Homi Bhabha National Institute, Dr. Ernest Borges Road, Parel, Mumbai, Maharashtra, India
2 Professor, Department of Medical Oncology, Tata Memorial Centre, Homi Bhabha National Institute, Dr Ernest Borges Road, Parel, Mumbai, Maharashtra, India
3 Professor, Department of Paediatric Oncology, Tata Memorial Centre, Homi Bhabha National Institute, Dr Ernest Borges Road, Parel, Mumbai, Maharashtra, India
4 Consultant in Medicine and Critical Care, PD Hinduja National Hospital, Mahim, Mumbai, Maharashtra, India
5 Consultant Microbiologist and Chair Infection Control, Hinduja Hospital, Mahim, Mumbai, Maharashtra, India
6 Consultant ID Physician, Jupiter Hospital, Pune, Deenanath Mangeshkar Hospital, Pune, Bharati Vidyapeeth, Deemed University Hospital, Pune, Courtesy Visiting Consultant, Hinduja Hospital Mumbai, Maharashtra, India
7 Professor and Head, Department of Pulmonary Medicine and Sleep Disorders, All India Institute of Medical Sciences, New Delhi, India
8 Sr Consultant, Pulmonary and Critical Care Medicine, Apollo Gleneagles Hospital, 58, Canal Circular Road, Kolkata, West Bengal, India
9 Head Department of Immunology, Prof of Pediatric Hematology and Oncology, Bai Jerbaiwadia Hospital for Children, Consultant, Hematologist, Nanavati Superspeciality Hospital, Director of Pediatric Hematology, Surya Hospitals, Mumbai, Maharashtra, India
10 Director, Intensive Care Unit, Fortis Hospital, Mulund Goregaon Link Road, Mulund (W), Mumbai, Maharashtra, India
11 Consultant and Head of Department, Critical Care Medicine, Ashoka - Medicover Hospital, Indira Nagar, Wadala Nashik, Maharashtra, India
12 Consultant Microbiologist, Microbiology Section, 5th Floor, S1 Building, PD Hinduja Hospital, Veer Savarkar Marg, Mahim, Mumbai, Maharashtra, India
13 Consultant Physician in Infectious Disease, Unison Medicare and Research Centre and Prince Aly Khan Hospital, Maharukh Mansion, Ali Bhai Premji Marg, Grant Road, Mumbai, Maharashtra, India
14 Associate Professor Intensive Care Medicine, Department of Anaesthesia, Critical Care and Pain, Tata Memorial Center, Homi Bhabha National Institute, Dr. E. Borges Road, Parel, Mumbai, Maharashtra, India
15 Associate Professor, Division of Critical Care Medicine, Department of Anaesthesia, Critical Care and Pain, Tata Memorial Center, Homi Bhabha National Institute, Dr. E. Borges Road, Parel, Mumbai, Maharashtra, India
16 Consultant in Critical Care, Director, ICU Sanjeevan and MJM Hospital, Pune, Maharashtra, India
17 Director and HOD, Neuro-Trauma Unit, Grant Medical Foundation, Ruby Hall Clinic, Pune, Maharashtra, India
18 Adjunct Professor - NBE, Chairman - Medanta Institute of Critical Care and Anesthesiology, Medanta The Medicity, Gurgaon, Haryana, India
19 Senior Resident, Division of Critical Care Medicine, Department of Anesthesiology, Critical Care and Pain, Tata Memorial Hospital, Homi Bhabha National Institute, Dr E Borges Road, Mumbai, Maharashtra, India
20 Specialist Senior Registrar, Department of Anaesthesia, Critical Care and Pain, Tata Memorial Center, HomiBhabha National Institute, Dr. E. Borges Road, Parel, Mumbai, Maharashtra, India
21 Specialist Senior Registrar, Department of Anaesthesia, Critical Care and Pain, Tata Memorial Center, Homi Bhabha National Institute, Dr. E. Borges Road, Parel, Mumbai, Maharashtra, India
22 Specialist Senior Registrar, Division of Critical Care Medicine, Department of Anaesthesia, Critical Care and Pain, Tata Memorial Center, Homi Bhabha National Institute, Dr. E. Borges Road, Parel, Mumbai, Maharashtra, India
23 Senior Registrar (Senior Resident), Division of Critical Care Medicine, Department of Anaesthesia, Critical Care and Pain, Tata Memorial Center, Homi Bhabha National Institute, Dr. E. Borges Road, Parel, Mumbai, Maharashtra, India
24 Senior Registrar, Division of Critical Care Medicine, Department of Anaesthesiology, Critical Care and Pain, Tata Memorial Center, Homi Bhabha National Institute, Dr. E. Borges Road, Parel, Mumbai, Maharashtra, India
25 Assistant Professor, Department of Medical Oncology, Tata Memorial Hospital, HomiBhabha National Institute, Mumbai, Maharashtra, India
26 Assistant Professor, Department of Pediatric Oncology, Tata Memorial Hospital, HomiBhabha National Institute, Dr E Borges Road, Mumbai, Maharashtra, India
27 Consultant, Critical Care, Fortis Hospital, 102, Nav Sai Shakti CHS, Near Bhoir Gymkhana, M Phule Road, Dombivali West Mumbai, Maharashtra, India
28 Junior Consultant, Critical Care Medicine, PD Hinduja National Hospital and MRC, Mumbai, Maharashtra, India
29 Professor and Head, Department of Anaesthesiology, Critical Care and Pain, Tata Memorial Hospital, Homi Bhabha National Institute, Dr. E. Borges Road, Parel, Mumbai, Maharashtra, India